



Review Article

Pharmacology for Better Drug Screening

Multi-Faceted Effects of Cannabis and Cannabinoids on the Male Reproductive System: A Review

Somenath Ghosh*

Assistant Professor, Post Graduate Department of Zoology, Rajendra College, Chapra-841301, Bihar, India.
E-mail: somenath9936439419@gmail.com

Abstract: *Cannabis sativa* is a hallucinating drug used in different parts of the world from ancient time. This is used as a part of various religious and social practices. Cannabis is a special type of Marijuana which can provide temporary relief from analgesia, body pain, and some other clinical conditions. Thus, importance of Cannabis is mostly described as a narcotic analgesic agent. But, the impacts of Cannabis on the reproductive health of females and males are multi-faceted and differentially fatal. Current researches suggest that cannabis may negatively impact male and female fertility conditions. In females, Cannabis can reduce female fertility by disrupting the hypothalamic release of gonadotropin-releasing hormone (GnRH), leading to reduced estrogen and progesterone production and an ovulatory menstrual cycles. In males, cannabis can cause changes in different ways in the reproductive system. At the tissue level, under chronic administration testicular morphology is changed which can exert its effect on sperm quality. Sperm quality is marked by different parameters like semen quality, sperm morphology, sperm mortality, and sperm motility. Upon chronic cannabis treatment, it was noted that almost all of the sperm parameters are significantly reduced. Reduced sperm parameters, eventually affect male reproductive hormones i.e. testosterone, Interstitial Cell Stimulating Hormone (ICSH), Luteinizing Hormone (LH), and Follicle Stimulating Hormone (FSH). These physiological and hormonal parameters finally cause reduced libido which may even lead to oligospermia and infertility. Till date, studies delineating cannabis effect on the male reproductive system is mostly centered on rodent models and it is mostly lacking in human and in sub-human primates. Considering the primate male reproductive system, these kinds of studies are haphazard and scanty. Thus, the lacunas of previous studies were identified and the present review focuses on the effects of cannabis on the male reproductive system in a systematic manner considering human and sub-human primates in particular.

Keywords: Cannabis, Effects, Hormones, Male, Marijuana, Reproduction, Reproductive potentiality, sperm parameters.

Article History	Date of Receiving 27 July 2020	Date of Revision 11 September 2020
	Date of Acceptance 16 September 2020	Date of Publishing 03 October 2020



*Corresponding Author

Dr. Somenath Ghosh, Assistant Professor, Post Graduate Department of Zoology, Rajendra College, Chapra-841301, Bihar, India.

Funding This research did not receive any specific grant from any funding agencies in the public, commercial or not for profit sectors.

Citation Dr. Somenath Ghosh, Multi-faceted effects of cannabis and cannabinoids on male reproductive system: A state of art review..(2020).Int J Pharm Sci. 11(4), P75-80 <http://dx.doi.org/10.22376/ijpbs.2020.11.4.p75-80>

This article is under the CC BY- NC-ND Licence (<https://creativecommons.org/licenses/by-nc-nd/4.0>)

Copyright © International Journal of Pharma and Bio Sciences, available at www.ijpbs.net

Int J Pharma Bio Sci., Volume 11., No 4 (Oct) 2020, pp P75-80



I. INTRODUCTION

Cannabis, which is a type of marijuana has been used by the people of the Indian subcontinent from time unknown for different social, ritual and entertainment purposes.¹ They not only use this herb as a part of the holy practice but also use it for recreational purposes². Irrespective of sex, this hallucinogenic agent is used by most parts of the world particularly by the populations of South America, India, Bangladesh, and Pakistan from a long time ago³. Reports suggesting the roles of *Cannabis* causing systemic neuropathy⁴, neuronal disability⁵, impaired fetal development⁶ and mal-functioning of male reproductive system⁷⁻¹⁰ are documented. But, no reports available depicting the effects of marijuana in the female reproductive system. The main causative agent of the effects of marijuana/cannabinoids is the endocannabinoid. This is a neutral lipid and highly conserved molecule throughout evolutionary history¹¹. They are having different derivatives like anandamide¹², 2-arachidonoylglycerol¹³ and Δ^9 -tetrahydrocannabinol (THC)¹⁴. However, among all of the fatty acid derivatives of cannabinoids or endocannabinoids (eCBs) the THC has now been established as the most important hallucinogenic agent of this molecule¹⁵. There is a literature suggesting the role of this THC in the regulation of functions of the central nervous system and thus regulating the reproductive functions by affecting/ modulating hypothalamic-pituitary-gonadal axis (HPG-axis)¹⁶ via its receptor CB1 and CB2¹⁷. Now it has been reported that CB1 receptors are localized mostly in the whole vertebrate central nervous system (CNS) and some peripheral tissues, whereas CB2 receptors are mostly expressed in peripheral tissues and immune cells, however, they have recently been found also in the CNS¹⁸. But, with all the advancement in psycho-neuro-endocrine research, to date, it is a matter of debate on how THC is going to regulate the reproductive system at the peripheral level. Some literatures suggest that there is a general agreement on the inhibitory effect exerted by cannabinoids and eCBs on GnRH release¹⁹ Thus, it is affecting the subsequent FSH and LH release and impairing both male and female reproduction²⁰. In the area of male fertility, cannabis use has been linked to reproductive hormone changes, altered semen parameters, and reduction in libido and sexual performance²¹. A detailed review of how marijuana affects male fertility at each point along the fertility axis is needed for clinicians to assess the potential risks that patients incur when using this substance. Although men are more likely to use drugs²² and have a substance use disorder²³, drug abuse can seriously impact even women's health. After alcohol and heroin, marijuana is the most common primary drug of abuse for women entering treatment for substance abuse²⁴. Females appear to be more sensitive to the behavioral and physiological effects of marijuana and marijuana-like substances²⁵, and treatment-seeking women endorse more severe marijuana withdrawal symptoms than treatment-seeking men²⁶. After tobacco and alcohol, marijuana is the most commonly abused substance by women of childbearing age²². According to the 2013 NSDUH, 5.4 percent of pregnant women and 11.4 percent of non-pregnant women ages 15 to 44 are currently illicit drug users²², with marijuana representing 64 to 79 percent of female drug use^{22, 27-29}. As marijuana becomes more widely legalized, marijuana used by women will likely increase²². Thus, with this brief introduction, it is evident that any form of marijuana can cause a detrimental effect on both male and female reproductive health. We have gone through several

literatures from Pubmed/Pubmed Central (PMC) and MedLine and with the results procured from our previous experiments on the impact of Cannabis on female reproduction [as published elsewhere]³⁰ the present article is designed. However, the present article is restricted to elucidate the roles of cannabis on male reproduction since the role of cannabis on female reproduction is a matter for another full-length chapter. This article may throw some light and put forward some recent knowledge on the impacts of marijuana and Cannabis in particular on reproductive impairment in males.

2. CANNABIS EFFECT ON MALE REPRODUCTION

Impacts of Cannabis consumption on male reproduction is multi-dimensional and can be exerted at many levels. Thus, it can be discussed under different sub headings.

2. 1. CANNABIS EFFECT ON SPERM PARAMETERS

2.1.1. SPERM COUNT AND CONCENTRATION

Cannabis use is strongly associated with reductions in sperm count and concentration in animal and human studies. Decreased epididymal sperm concentrations were observed in mature male rats exposed to 16 puffs per day of marijuana, comparable to the recreational level in humans, for 75 days³¹. This effect was replicated in a study, in which, 3 to 6 mg/kg of the *Cannabis sativa* derivative bhang was administered in adult male mice. The results of the study demonstrated a significantly depressed sperm count³². Human studies have shown similar findings. A Danish cohort study on marijuana use in 1,215 participants revealed similar changes³³. Men who reported using marijuana more than once per week had a 28% lower sperm concentration and a 29% lower sperm count than men who had never used marijuana. In a study in which 16 chronic marijuana smokers were exposed to 4 weeks of high dose marijuana the time to a reduced sperm count was 5 to 6 weeks after initiating marijuana use³⁴. In human and animal models cannabis has shown strong links to reduced sperm count and concentration which may be linked to arrested spermatogenesis. Future work is needed to elucidate causal mechanisms.

2.1.2. MORPHOLOGY

Cannabis use also appears to induce considerable morphological changes in sperm. In a 1978 study, Zimmerman *et al* treated male mice for 5 consecutive days with intraperitoneal injections of the marijuana components THC, cannabinoid, or cannabiol³⁵. On microscopy, 35 days after exposure mice treated with THC and cannabiol had a significantly higher incidence of abnormal morphology than the control group, such as banana-shaped, amorphous, folded, or hookless heads. Despite morphological changes the research suggests that cannabis does not induce chromosome breakage in sperm. Generoso *et al*³⁶ administered 50 mg/kg THC 5 times per week for 6 weeks in 498 male mice³⁶. After mating them with females no increase was observed in fetal dominant lethal mutations or heritable translocations over those in controls. These findings were supported by a study by Berryman *et al*, who found no THC induced increase in preimplantation loss, fetal mortality, or the mutation index in fetuses fathered by male mice chronically dosed with THC³⁷. In animal and human models

evidence suggests that cannabis induces morphological changes in sperm while genetic material is preserved.

2.1.3. MOTILITY AND ENERGY METABOLISM

The most extensive body of evidence for cannabis related alterations to sperm is for sperm motility. Whan *et al*³⁸ noted spermatotoxic effects of THC by incubating sperm with THC at therapeutic doses, similar to concentrations shown to relieve pain or reduce spasticity in humans with multiple sclerosis (0.032 mM) and recreational concentrations (4.8 and 0.32 mM) for 3 hours, and then measuring motility by computer-assisted semen analysis³⁸. Therapeutic and recreational THC concentrations also resulted in reduced straight-line velocity. A similar decrease in sperm motility was seen in an analysis of semen samples from 16 healthy, chronic marijuana users after 4 weeks of high dose marijuana³⁴. Barbonetti *et al* elucidated the mechanism of these findings by establishing a link between CBI and sperm mitochondrial activity³⁹. In sperm incubated with the CBI receptor agonist Met-AEA a significant reduction was observed in mitochondrial trans-membrane potential. When the sperm was placed under glycolysis blockade, causing them to switch oxidative phosphorylation, the introduction of Met-AEA abolished sperm motility. The link between cannabis and sperm mitochondrial dysfunction was furthered by Badawy *et al*⁴⁰, who added THC to semen and measured the oxygen concentration as a marker of respiration. Upon the addition of THC mitochondrial respiration immediately declined and was concentration-dependent in effect. The results were much more pronounced in washed sperm than in neat semen, suggesting that seminal plasma contains some protective factors. These various investigations suggest that through the action of cannabis on the CBI receptor the mitochondrial activity of sperm is reduced and as a result motility is significantly impaired. Although Met-AEA and THC administration in the laboratory has helped map potential pathways, to our knowledge it is not known whether these effects are fully replicated in the male testes. Future testing should be done to explore whether mitochondrial impairment is present in the semen of chronic cannabis users.

2.1.4. VIABILITY

Cannabis also has a detrimental effect on sperm viability. Rossato *et al*⁴¹ incubated semen samples with the endocannabinoid AEA at varying concentrations and found that viability was decreased in a manner at supraphysiological AEA concentrations. Reduced sperm viability related to cannabis has also been investigated using the highly specific CBI receptor antagonist rimonabant (SRI141716). Cobelliset *al*⁴² found that adding a micromolar concentration of rimonabant induced a small but significant increase in the number of viable spermatozoa. Aquila *et al*⁴³ reported similar findings with 1 and 10 nM concentrations of rimonabant increasing sperm viability with no further viability changes observed at higher concentrations. While the cannabinoid system has clear links to sperm viability, future work should be done to confirm these findings with exogenous cannabinoids as well as in the *in vivo* setting.

2.1.5. FERTILIZATION CAPACITY

Research suggests that the cannabinoid signaling pathway may be involved in inhibiting sperm capacitation and activation.

Using high performance liquid chromatography Schuelet *al*⁴⁴ observed that high levels of AEA are present in seminal plasma and in progressively decreasing amounts in oviductal and follicular fluid, indicating that sperm are exposed to progressively decreasing AEA levels along the entire fertilization path⁴⁵. The authors speculated that high AEA levels maintain sperm in a quiescent state and the decrease in AEA levels which occurs in the fertilization environment enables sperm to become activated. These data suggest that increase in cannabinoid levels may interfere with sperm activation and may be especially pertinent in the female reproductive tract, where the sperm depend on for tightly regulated AEA levels to maintain proper function. Rossato *et al* reported that AEA inhibits the capacitation induced acrosome reaction of human sperm after incubation in capacitating medium⁴¹. Using boar sperm Maccarrone *et al* found that Met-AEA reduced sperm capacitation in a time dependent manner⁴⁶. They also noted that this effect was mediated by the CBI receptor since when rimonabant, which blocks CBI, was added, Met-AEA produced no change in capacitation. Schuelet *al*⁴⁴ used the cannabinoid agonist AM-365 to identify concentration-dependent stimulation and inhibition of sperm hyperactivated motility, which is a state needed for sperm to reach the egg surface and which assists with penetration of the zonapellucida. Current work suggests that the endocannabinoid system is intimately linked to the fertilization process in the male and female reproductive tracts. Given the well described inhibitory effects, cannabis is likely to have negative impacts on fertilizing potential.

3. CANNABIS EFFECTS ON REPRODUCTIVE HORMONES:

3.1. FOLLICLE STIMULATING HORMONE

Relatively few studies have focused on cannabis and FSH levels, and most have observed no effect. Cone *et al* found no significant change in FSH levels in 4 healthy males with a history of frequent marijuana use before and after 2 marijuana cigarettes per day for 3 consecutive days⁴⁷. Vescovi *et al* observed that cannabis did not alter the response of FSH to gonadotropin-releasing hormone in 10 male chronic marijuana users given gonadotropin-releasing hormone intravenously⁴⁸. A depression in FSH levels was observed only by Kolodny *et al*, who compared plasma hormone levels among 11 men who used 5 to 9 marijuana cigarettes per week, 9 who used 10 or more marijuana cigarettes per week, and normal controls²⁹. Based on current studies, FSH may not be affected by cannabis except perhaps in the limited case of heavy chronic use. Human studies to date have been limited in suggestive power due to the small cohort sizes, leaving considerable room for further validation with larger sample size investigations.

3.2. LUTEINIZING HORMONE

In human and animal models LH is consistently lowered by cannabis^{49, 50}. In the single exception, Kolodny *et al*²⁹ did not observe any significant difference in plasma LH levels between men who smoked 5 to 9 marijuana cigarettes per week and men who smoked 10 or more per week. However, the variation in marijuana use levels in this study may have been insufficient to induce LH variations. The relationship between cannabis and LH was strengthened in a study by Wenger *et al*, who used polyclonal antibodies against CBI and CB2 to localize individual cells expressing

cannabinoid receptors⁵¹. The CBI receptor was found in the anterior pituitary in LH secreting gonadotrophic cells. Wenger *et al* reaffirmed these results after administering AEA to wild-type and CBI knockout mice, which revealed decreased LH secretion in the wildtype mice but unchanged LH levels in the CBI knockout mice⁵². As is the case with FSH related investigations, understanding how cannabis impacts LH would be improved by larger randomized, controlled trials in human subjects.

3.3. TESTOSTERONE

The reported effect of cannabis on serum testosterone levels is widely variable across current studies. In early work in 20 chronic marijuana users, Kolodny *et al* found a significant reduction in testosterone levels between chronic and never marijuana users²⁹. The evidence that cannabis depresses testosterone levels relies heavily on animal studies. In contrast to findings in animals, most human studies support the conclusion that testosterone levels are not significantly changed by cannabis use. A 1974 study by Mendelson *et al* in 27 chronic marijuana users who were administered marijuana for 21 days showed no significant changes in plasma testosterone levels⁵². In a 1986 study of 4 male chronic marijuana users given 2 marijuana cigarettes per day depressed free testosterone levels were observed but the levels did not significantly differ from baseline⁴⁷. In a later study, free testosterone levels were compared in 41 normal controls, and in 66 Pakistani men who smoked cannabis daily or regularly consumed cannabis tea³⁵. No significant difference was observed in plasma testosterone levels between the cannabis users and the normal controls. Although the sample size was limited in these early human studies, they suggest that cannabis consumption does not significantly alter testosterone levels. It is only recently that large cohort studies of cannabis users have been possible. To date, these studies have continued the trend of presenting conflicting or inconclusive evidence on the link between cannabis use and testosterone levels. The first large cohort study on the effects of cannabis use was performed by Gundersen *et al* in 2015 using a registry of 1,215 Danish men undergoing a compulsory medical examination to determine fitness for service³³. Testosterone levels were 7% higher in self-reported marijuana smokers than in nonusers. This was within the same range of testosterone elevation observed in cigarette smokers in the cohort. The authors cautioned that increased testosterone levels in marijuana users could not be separated from the effect of tobacco smoking alone. A second major cohort study was done in 2017 by Thistle *et al*⁵³. They used data on 1,577 American men using data from the 2011 to 2012 United States National Health and Nutrition

6. REFERENCES

- Ashton CH, Moore PB, Gallagher P, Young AH. Cannabinoids in bipolar affective disorder: a review and discussion of their therapeutic potential. *J Psychopharmacol.* 2005;19(3):293-300. doi: 10.1177/0269881105051541, PMID 15888515.
- Aversa A, Rossi F, Francomano D, Bruzziches R, Bertone C, Santemma V, Spera G. Early endothelial dysfunction as a marker of vasculogenic erectile dysfunction in young habitual cannabis users. *Int J Impot Res.* 2008;20(6):566-73. doi: 10.1038/ijir.2008.43, PMID 18997809.

Survey with several novel outcomes. No difference was observed in serum testosterone levels between ever and never users of marijuana. However, serum testosterone levels showed an inverse association with time since the last regular use of marijuana, and since the last marijuana use. This indicated that decency rather than frequency of use may have the strongest relationship with serum testosterone levels. Additional large, population-based samples are needed to clarify currently conflicting reported effects of cannabis on testosterone levels.

4. CONCLUSION

As cannabis increasingly gains legalized status across the United States, the popularity and prevalence of use continue to grow. Although medically it demonstrates therapeutic promise in some areas such as multiple sclerosis and chronic neuropathic pain, the potential adverse effects remain widely under studied. Current research shows that cannabis likely has negative impacts at several points along the male fertility pathway. Human sperm express cannabinoid receptors, suggesting that they are directly impacted by alterations in the balance of the end cannabinoid system. The effect of cannabis on testosterone levels is largely undetermined while LH levels appear to be lowered and FSH levels are unchanged. The strongest evidence for the deleterious effects of cannabis on male reproductive capacity is its impact on semen parameters. Studies demonstrate reduced sperm count and concentration, morphological changes, reduced motility and viability, and decreased fertilizing capacity in animals and humans exposed to marijuana or cannabis derivatives. Furthermore, animal studies suggest that cannabis has a role in testicular atrophy. While cannabis may increase libido in the short term, chronic use may diminish erectile function in men. The evidence presented to date largely depends on animal models, *in vitro* studies of endogenous cannabinoid compounds and retrospective analyses. The ethical and legal complications of an *in vivo*, controlled study drive the limited amount of data presented in human subjects, a limitation which is unlikely to abate going forward. Future studies should focus on gathering large cohort data in national surveys, similar to the voluntary, cannabis related data collection done with compulsory military fitness examinations in different countries. These types of studies are needed to confirm that animal models can be translated into human experience.

5. CONFLICT OF INTEREST

Conflict of interest declared none.

- Battista N, Pasquariello N, Di Tommaso MM, Maccarrone M. Interplay between endocannabinoids, steroids and cytokines in the control of human reproduction. *J Neuroendocrinol.* 2008;20;Suppl 1:82-9. doi: 10.1111/j.1365-2826.2008.01684.x, PMID 18426505.
- Bayewitch M, Rhee MH, Avidor-Reiss T, Breuer A, Mechoulam R, Vogel Z. (-)-Delta9-tetrahydrocannabinol antagonizes the peripheral cannabinoid receptor-mediated inhibition of adenylyl cyclase. *J Biol Chem.* 1996; 271(17):9902-5.

- doi: 10.1074/jbc.271.17.9902, PMID 8626625.
5. Brown TT, Dobs AS. Endocrine effects of marijuana. *J Clin Pharmacol.* 2002;42(S1);Suppl 1 [Suppl 90:90S-6S]:90S-6S. doi: 10.1002/j.1552-4604.2002.tb06008.x, PMID 12412841.
 6. Cacciola G, Chioccarelli T, Mackie K, Meccariello R, Ledent C, Fasano S, Pierantoni R, Cobellis G. Expression of type-I cannabinoid receptor during rat postnatal testicular development: possible involvement in adult Leydig cell differentiation. *Biol Reprod.* 2008;79(4):758-65. doi: 10.1095/biolreprod.108.070128, PMID 18614700.
 7. Cobellis G, Cacciola G, Scarpa D, Meccariello R, Chianese R, Franzoni MF, Mackie K, Pierantoni R, Fasano S. Endocannabinoid system in frog and rodent testis: type-I cannabinoid receptor and fatty acid amide hydrolase activity in male germ cells. *Biol Reprod.* 2006;75(1):82-9. doi: 10.1095/biolreprod.106.051730, PMID 16611862.
 8. Chakravarty I, Shah PG, Sheth AR, Ghosh JJ. Mode of action of delta-9-tetrahydrocannabinol on hypothalamo-pituitary function in adult female rats. *J Reprod Fertil.* 1979;57(1):113-5. doi: 10.1530/jrf.0.0570113, PMID 390133.
 9. Cravatt BF, Giang DK, Mayfield SP, Boger DL, Lerner RA, Gilula NB. Molecular characterization of an enzyme that degrades neuromodulatory fatty-acid amides. *Nature.* 1996;384(6604):83-7. doi: 10.1038/384083a0, PMID 8900284.
 10. Campbell VA. Tetrahydrocannabinol-induced apoptosis of cultured cortical neurones is associated with cytochrome c release and caspase-3 activation. *Neuropharmacology.* 2001; 40(5): 702-9. doi: 10.1016/s0028-3908(00)00210-0, PMID 11311898.
 11. Dalterio S, Bartke A, Burstein S. Cannabinoids inhibit testosterone secretion by mouse testes *in vitro*. *Science.* 1977;196(4297):1472-3. doi: 10.1126/science.867048, PMID 867048.
 12. Singh SK, Chakravarty S. Antispermatozoic and antifertility effects of 20, 25-diazacholesterol dihydrochloride in mice. *Reprod Toxicol.* 2003;17(1):37-44. doi: 10.1016/s0890-6238(02)00075-8, PMID 12507656.
 13. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, Gibson D, Mandelbaum A, Etinger A, Mechoulam R. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science.* 1992;258(5090):1946-9. doi: 10.1126/science.1470919, PMID 1470919.
 14. Dhawan K, Kumar S, Sharma A. Reversal of cannabinoids (delta9-THC) by the benzoflavone moiety from methanol extract of *Passiflora incarnata* Linneaus in mice: a possible therapy for cannabinoid addiction. *J Pharm Pharmacol.* 2002;54(6):875-81. doi: 10.1211/0022357021779069, PMID 12079005.
 15. Dixit VP, Lohiya NK. Effects of cannabis extract on the response of accessory sex organs of adult male mice to testosterone. *Indian J Physiol Pharmacol.* 1975;19(2):98-100. PMID 1158438.
 16. Dixit VP, Gupta CL, Agrawal M. Testicular degeneration and necrosis induced by chronic administration of cannabis extract in dogs. *Endokrinologie.* 1977;69(3):299-305. PMID 913356.
 17. EL-Gohary M, Eid MA. Effect of cannabinoid ingestion (in the form of bhang) on the immune system of high school and university students. *Hum Exp Toxicol.* 2004;23(3):149-56. doi: 10.1191/0960327104ht426oa, PMID 15119535.
 18. Gammon CM, Freeman Jr GM, Xie W, Petersen SL, Wetsel WC. Regulation of gonadotropin-releasing hormone secretion by cannabinoids. *Endocrinology.* 2005;146(10):4491-9. doi: 10.1210/en.2004-1672, PMID 16020480.
 19. Gaoni Y, Mechoulam R. Isolation, structure, and partial synthesis of an active constituent of hashish. *J Am Chem Soc.* 1964;86(8):1646-7. doi: 10.1021/ja01062a046.
 20. Gye MC, Kang HH, Kang HJ. Expression of cannabinoid receptor 1 in mouse testes. *Arch Androl.* 2005;51(3):247-55. doi: 10.1080/014850190898845, PMID 16025865.
 21. Bari M, Battista N, Pirazzi V, Maccarrone M. The manifold actions of endocannabinoids on female and male reproductive events. *Front Biosci (Landmark Ed).* 2011;16:498-516. doi: 10.2741/3701, PMID 21196184.
 22. Substance Abuse and Mental Health Services Administration. Results from the 2013 national survey on drug use and health: summary of national findings. Rockville, MD: Substance Abuse and Mental Health Services Administration; 2014. Report No. : NSDUH Series H-48. HHS. Publication no. SBE OFFICE of multidisciplinary activities. p. 14-4863 [internet] [cited 11/9/2020]. Available from: <http://www.samhsa.gov/data/sites/default/files/NSDUHresultsPDFwHTML2013/web/NSDUHresults2013.pdf>.
 23. Grant BF, Saha TD, Ruan WJ, Goldstein RB, Chou SP, Jung J, Zhang H, Smith SM, Pickering RP, Huang B, Hasin DS. Epidemiology of DSM-5 drug use disorder: results from the national epidemiologic survey on alcohol and related conditions-III. *JAMA Psychiatry.* 2016;73(1):39-47. doi: 10.1001/jamapsychiatry.2015.2132,
 24. Substance Abuse and Mental Health Services Administration, Center for Behavioral Health Statistics and Quality. Treatment episode data set (TEDS). Available from: https://www.samhsa.gov/data/sites/default/files/TEDS2012N_Web.pdf. National admission to substance abuse treatment services. Rockville, MD: SUBSTANCE ABUSE and mental health services administration; 2002-2012. p. 2014. Report No. : BHSiS Series S- 71, HHS Publication No. (SMA) 14-4850.
 25. Craft RM. Sex differences in behavioral effects of cannabinoids. *Life Sci.* 2005;77(20):2471-8. doi: 10.1016/j.lfs.2005.04.019, PMID 15958268.
 26. Herrmann ES, Weerts EM, Vandrey R. Sex differences in cannabis withdrawal symptoms among treatment-seeking cannabis users. *Exp Clin Psychopharmacol.* 2015;23(6):415-21. doi: 10.1037/pha0000053, PMID 26461168.
 27. Sherwood RA, Keating J, Kavvadia V, Greenough A, Peters TJ. Substance misuse in early pregnancy and relationship to fetal outcome. *Eur J Pediatr.* 1999;158(6):488-92. doi: 10.1007/s004310051126, PMID 10378398.
 28. García-Serra J, Ramis J, Simó S, Joya X, Pichini S, vall O, García-Algar O. Alternative biological materials to detect prenatal exposure to drugs of abuse in the third trimester of pregnancy. *Ann Pediatr (Barc).* 2012;77(5):323-8. doi: 10.1016/j.anpedi.2012.02.019, PMID 22513392.

29. Ebrahim SH, Gfroerer J. Pregnancy-related substance use in the united states during 1996-1998. *Obstet Gynecol.* 2003;101(2):374-9. doi: 10.1016/s0029-7844(02)02588-7, PMID 12576263.
30. Ghosh S, Rai SK. Chronic cannabis induced oxidative stress and reproductive containment in female mice. *Int J Green Pharm.* 2018;12:227. doi: 10.22377/ijgp.v12i03.1956.
31. Huang HF, Nahas GG, Hembree WC 3rd. Effects of marihuana inhalation on spermatogenesis of the rat. *Adv Biosci.* 1978;22-23:419-27. doi: 10.1016/b978-0-08-023759-6.50037-8, PMID 756840.
32. Banerjee A, Singh A, Srivastava P, Turner H, Krishna A. Effects of chronic bhang (cannabis) administration on the reproductive system of male mice. *Birth Defects Res B Dev Reprod Toxicol.* 2011;92(3):195-205. doi: 10.1002/bdrb.20295, PMID 21678546.
33. Gundersen TD, Jørgensen N, Andersson AM, Bang AK, Nordkap L, Skakkebaek NE, Priskorn L, Juul A, Jensen TK. Association between use of marijuana and male reproductive hormones and semen quality: a study among 1,215 healthy young men. *Am J Epidemiol.* 2015;182(6):473-81. doi: 10.1093/aje/kwv135, PMID 26283092.
34. Hembree WC 3rd, Nahas GG, Zeidenberg P, Huang HF. Changes in human spermatozoa associated with high dose marihuana smoking. *Adv Biosci.* 1978;22-23:429-39. doi: 10.1016/b978-0-08-023759-6.50038-x, PMID 574469.
35. Zimmerman AM, Zimmerman S, Raj AY. Effects of cannabinoids on spermatogenesis in mice. *Adv Biosci.* 1978;22-23:407-18. doi: 10.1016/b978-0-08-023759-6.50036-6, PMID 756839.
36. Generoso WM, Cain KT, Cornett CV, Shelby MD. Tests for induction of dominant-lethal mutations and heritable translocations with tetrahydrocannabinol in male mice. *Mutat Res.* 1985;143(1-2):51-3. doi: 10.1016/0165-7992(85)90104-6, PMID 2987686.
37. Berryman SH, Anderson RA Jr, Weis J, Bartke A. Evaluation of the co-mutagenicity of ethanol and delta 9-tetrahydrocannabinol with Trenimon. *Mutat Res.* 1992;278(1):47-60. doi: 10.1016/0165-1218(92)90285-8, PMID 1370119.
38. Whan LB, West MC, McClure N, Lewis SE. Effects of delta-9-tetrahydrocannabinol, the primary psychoactive cannabinoid in marijuana, on human sperm function in vitro. *Fertil Steril.* 2006;85(3):653-60. doi: 10.1016/j.fertnstert.2005.08.027, PMID 16500334.
39. Barbonetti A, Vassallo MR, Fortunato D, Francavilla S, Maccarrone M, Francavilla F. Energetic metabolism and human sperm motility: impact of CB₁ receptor activation. *Endocrinology.* 2010;151(12):5882-92. doi: 10.1210/en.2010-0484, PMID 20962050.
40. Badawy ZS, Chohan KR, Whyte DA, Penefsky HS, Brown OM, Souid AK. Cannabinoids inhibit the respiration of human sperm. *Fertil Steril.* 2009;91(6):2471-6. doi: 10.1016/j.fertnstert.2008.03.075, PMID 18565513.
41. Rossato M, Ion Popa F, Ferigo M, Clari G, Foresta C. Human sperm express cannabinoid receptor Cb1, the activation of which inhibits motility, acrosome reaction, and mitochondrial function. *J Clin Endocrinol Metab.* 2005;90(2):984-91. doi: 10.1210/jc.2004-1287, PMID 15562018.
42. Chianese R, Ciaramella V, Scarpa D, Fasano S, Pierantoni R, Meccariello R. Endocannabinoids and Endovanilloids: A Possible Balance in the Regulation of the Testicular GnRH Signalling. *Int J Endocrinol.* Volume 2013, Article ID 904748, 9 pages. doi: 10.1155/2013/904748.
43. Aquila S, Guido C, Santoro A, Perrotta I, Laezza C, Bifulco M, Sebastiano A. Human sperm anatomy: ultrastructural localization of the cannabinoid I receptor and a potential role of anandamide in sperm survival and acrosome reaction. *Anat Rec (Hoboken).* 2010;293(2):298-309. doi: 10.1002/ar.21042, PMID 19938110.
44. Schuel H, Burkman LJ. A tale of two cells: endocannabinoid-signaling regulates functions of neurons and sperm. *Biol Reprod.* 2005;73(6):1078-86. doi: 10.1095/biolreprod.105.043273, PMID 16120829.
45. Schuel H, Burkman LJ, Lippes J, Crickard K, Mahony MC, Giuffrida A, Picone RP, Makriyannis A. Evidence that anandamide-signaling regulates human sperm functions required for fertilization. *Mol Reprod Dev.* 2002;63(3):376-87. doi: 10.1002/mrd.90021, PMID 12237954.
46. Maccarrone M, Barboni B, Paradisi A, Bernabò N, Gasperi V, Pistilli MG, Fezza F, Lucidi P, Mattioli M. Characterization of the endocannabinoid system in boar spermatozoa and implications for sperm capacitation and acrosome reaction. *J Cell Sci.* 2005;118(19):4393-404. doi: 10.1242/jcs.02536, PMID 16144868.
47. Cone EJ, Johnson RE, Moore JD, Roache JD. Acute effects of smoking marijuana on hormones, subjective effects and performance in male human subjects. *Pharmacol Biochem Behav.* 1986;24(6):1749-54. doi: 10.1016/0091-3057(86)90515-0, PMID 3016764.
48. Wenger T, Rettori V, Snyder GD, Dalterio S, McCann SM. Effects of delta-9-tetrahydrocannabinol on the hypothalamic-pituitary control of luteinizing hormone and follicle-stimulating hormone secretion in adult male rats. *Neuroendocrinology.* 1987;46(6):488-93. doi: 10.1159/000124870, PMID 2827048.
49. Vescovi PP, Pedrazzoni M, Michelini M, Maninetti L, Bernardelli F, Passeri M. Chronic effects of marihuana smoking on luteinizing hormone, follicle-stimulating hormone and prolactin levels in human males. *Drug Alcohol Depend.* 1992;30(1):59-63. doi: 10.1016/0376-8716(92)90036-c, PMID 1591981.
50. Martín-Calderón JL, Muñoz RM, Villanúa MA, del Arco I, Moreno JL, de Fonseca FR, Navarro M. Characterization of the acute endocrine actions of (-)-11-hydroxy-delta8-tetrahydrocannabinol-dimethylheptyl (HU-210), a potent synthetic cannabinoid in rats, [De Fonseca FR, Navarro M]. *Eur J Pharmacol.* 1998;344(1):77-86. doi: 10.1016/S0014-2999(97)01560-4, PMID 9580419.
51. Wenger T, Fernández-Ruiz JJ, Ramos JA. Immunocytochemical demonstration of CB1 cannabinoid receptors in the anterior lobe of the pituitary gland. *J Neuroendocrinol.* 1999;11(11):873-8. doi: 10.1046/j.1365-2826.1999.00402.x,
52. Mendelson JH, Kuehnle J, Ellingboe J, Babor TF. Plasma testosterone levels before, during and after chronic marihuana smoking. *N Engl J Med.* 1974;291(20):1051-5. doi: 10.1056/NEJM197411142912003, PMID 4415097.
53. Thistle JE, Graubard BI, Braunlin M, Vesper H, Trabert B, Cook MB, McGlynn KA. Marijuana use and serum testosterone concentrations among U.S. males. *Andrology.* 2017;5(4):732-8. doi: 10.1111/andr.12358, PMID 28395129.