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ANTIBACTERIAL ACTIVITIES OF METABOLITES FROM CORYNEBACTERIUM SPP. STRAINS ISOLATED FROM THE REPRODUCTIVE TRACT OF A HEALTHY WOMAN AGAINST HUMAN PATHOGENIC BACTERIA

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ABSTRACT

The genus *Corynebacterium* spp., also known as "diphtheroids" or "coryneform" includes diverse group of microorganisms inhabiting different biotopes of the human body. In this work we have studied the influence of metabolites from *Corynebacterium* spp. strains isolated from the reproductive tract of a healthy woman on growth, biofilm formation and on developed biofilms (24h grown) *K. pneumoniae, E. coli, P. aeruginosa* and *S. aureus*. We found that all human bacterial pathogens showed slower growth in the presence of metabolites of *Corynebacterium*. All the isolated *Corynebacterium* strains to inhibit biofilm formation and destroy developed biofilms (24h grown) *K. pneumoniae, E. coli, P. aeruginosa* and *S. aureus*. The obtained data reveals the important role of *Corynebacteria* in protecting the vaginal biotope from infection in those women who do not have *Lactobacilli* and suggest a probiotic potential of these *Corynebacterium* strains as an anti-bacterial agent in the vagina.

KEYWORDS: Corynebacterium, antibacterial activity, antibiofilm activity, pathogenic bacteria





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INTRODUCTION

The genus Corynebacterium belongs to the family Corynebacteriaceae of the phylum Actinobacteria and includes straight or slightly curved rod-shaped, Grampositive, catalase-positive, non-spore-forming, nonmicro(aerobic) microorganisms.1 motile Among Corynebacteria, the most significant human pathogen diphtheria worldwide.² In addition In addition, there nondiphtheriae Corynebacteria, major components of the normal flora of human skin and mucous membranes. are commonly isolated from clinical specimens. It is widely known that nondiphtheriae Corynebacteria associated with cases of endocarditis, meningitis, arthritis, sinusitis, skin wounds and intrauterine infections in both immunocompromised immunocompetent patients.³ At the same time, several authors have described the important role the participation of particular strains of nondiphtheriae Corynebacteria in the protection of the different biotopes of the human body from infections. Uehara et al., reported the elimination of nasal Staphylococcus aureus by application of Corynebacterium sp.4 Bomar et al., showed that Corynebacterium accolens, inhibits growth of Streptococcus pneumoniae by releasing antibacterial free fatty acids from host skin surface triacylglycerols.5 Wysocki et al. demonstrated that lipophilic species of Corynebacterium inhabiting skin as residents produces substances that can regulate the composition of natural flora. In the course of research an inhibiting substance (BLIS) was isolated with its evident effect on S. aureus, S. epidermidis, C. diphtheriae and Propionibacterium spp.6 Matthew M. Ramsey et al. found that S. aureus interactions with Corynebacterium spp. diminish S. aureus virulence.7 In vaginal biotope nondiphtheriae Corynebacteria are found in women regardless of age and microecological status of the biotope. 8,9,10 In girls before puberty nondiphtheriae Corynebacteria along with Staphylococcus epidermidis constitute the main part – 80% of the vaginal microbiota. 11 Their number also is increased in pregnant and postpartum women. 12 Previously, we carried out a pilot study on intermicrobial interaction Corynebacteria and Lactobacilli isolated from the reproductive tract of women. We found that the metabolites Corynebacteria greatly increased antagonistic activity of the peroxide producing Lactobacilli against pathogenic and opportunistic microorganisms. 13 These results show an important role nondiphtheriae Corynebacteria in the protection of the female reproductive tract against infection. However, research on their own antagonistic activity of Corynebacteria isolated from the reproductive tract of women to pathogenic and opportunistic microorganisms has not yet been conducted. Therefore, the aim of the current study is to determine the antibacterial activity of metabolites from Corynebacterium spp. against four important human bacterial pathogens.

MATERIALS AND METHODS

Isolation and identification of Corynebacterium

This study was conducted in the Laboratory of study mechanism formation microbiocenosis of human,

Institute for Cellular and Intracellular Symbiosis UrB RAS, Orenburg, Russia. The four *Corynebacterium* strains used in this study were previously isolated from vaginal smears of healthy women and were previously characterized by their properties increased the antagonistic activity of peroxide producing *Lactobacilli*. These strains are *C. amycolatum ICIS 9, C. amycolatum ICIS 53, C. minutissimum ICIS 5 and C. xerosis ICIS 99. ¹³ Corynebacteria were identified on a set of morphological, cultural and biochemical properties (API Coryne 2.0 biochemical identification system (bioMérieux, France) according to manufacturer's instructions).*

Bacterial strains

Four human bacterial pathogens obtained from microbial culture collection, Institute for Cellular and Intracellular Symbiosis UrB RAS, Orenburg, Russia. These strains are *Klebsiella pneumoniae, Escherichia coli, Pseudomonas aeruginosa* and *Staphylococcus aureus*. For the experiment, the strains were prepared according to Prabakaran and Kalimuthu (2013). ¹⁴

Preparation of metabolites

Corynebacterium strains was grown in tryptic soy broth (TSB) for 24 h at 37°C. Metabolite was obtained by centrifuging the culture (10000×g, 20 min, 4°C), purified by filtration (0.22 μ m, Millipore Nihon, USA).

Inhibitory effect of metabolites from Corynebacterium on human bacterial pathogens

The direct antagonism metabolites Corynebacterium strains against human bacterial pathogens was monitored by turbidimetry. Aliquot of 100 μL of preinoculated pathogens (about 10⁶ CFU/ml) in TSB broth was transferred to each well of 96 well polystyrene plates, the same volume of metabolites from Corynebacterium strains added to each well in replica and incubated at 37°C for 7h. Control wells contained culture medium and the tested strain without adding metabolites. Samples were taken at intervals of 1 hour, and their optical density (OD) was determined using a microplate photometer Thermo Scientific Multiskan FC at 600 nm.

Inhibitory effects of metabolites from Corynebacterium on biofilm formation of human bacterial pathogens

Inhibitory effect of metabolites from Corynebacterium strains isolates against biofilm formation by human bacterial pathogens was determined using the commonly used 96 wells polystyrene plate method and was performed according to O'Toole and Kolter (1998)¹ with slight modifications. Aliquot of 100 µL of preinoculated pathogens (about 10⁶ CFU/ml) in TSB broth was transferred to each well of 96 well plates, the same volume of metabolites from Corynebacterium strains added to each well. Control wells contained culture medium and the tested strain without adding metabolites. Further, the experiment was performed according to the procedure described by Wojnicz and Tichaczek-Goska (2013)¹⁶. After incubation at 37°C for 24 h the wells were rinsed thoroughly with phosphate buffered saline (PBS) in order to remove nonadherent bacteria. Bacterial cells bound to the walls of the wells were stained with 1% (w/v) crystal violet (Sigma) for 15

min, and then rinsed thoroughly with PBS. Afterwards the dye bound to the adherent bacterial cells was resolubilized with 95% (v/v) ethanol. The OD of each well was measured at 570 nm using a microplate photometer. The results were expressed as percentage of biofilm inhibition according to the following equation:

Percentage inhibition = (A1-A2)/A1×100

where A1— the absorbance of the control group (without bacterial metabolites).

A2— the absorbance of the treatment group (with media containing bacterial metabolites).

The influence of metabolites from Corynebacterium strains on pre-formed biofilms

The influence of metabolites from *Corynebacterium* strains on pre-formed biofilms quantified according to the procedure described by Dalili *et al.*, (2015) 17 with slight modifications. Aliquot of 200 μL of bacterial pathogens (about 10^6 CFU/ml) in TSB broth was transferred to wells of 96 well plates. After incubation for 24h, the media were poured out from the wells and washed with sterile PBS buffer. Then, 200 μL of metabolites was added to each well following the incubation for 24 h. The culture without metabolites was served as control. The percentage of biofilms inhibition at metabolites for each microorganism was calculated according to the above equation.

STATISTICAL ANALYSIS

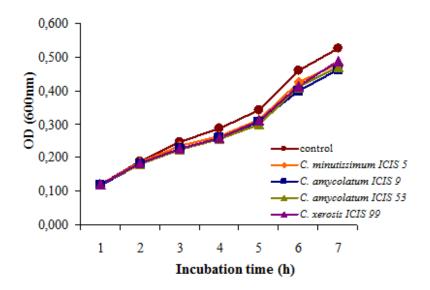
Statistical analysis was done using the statistical software "STATISTICA 10.0". Differences were considered significant when P<0.05.

RESULTS

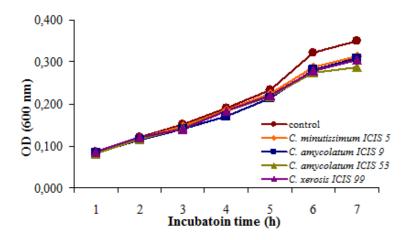
Four Corynebacterium strains previously were isolated from the reproductive tract of healthy women and were analyzed with regard to their morphology, cultural and biochemical characteristics. All strains were Grampositive, catalase-positive, non-spore-forming, non-motile straight or slightly curved rod-shaped cells. The identification of Corynebacterium isolates was confirmed by determining the biochemical properties (API Coryne 2.0 biochemical system (bioMérieux, France) according to manufacturer's instructions. These strains are C. amycolatum ICIS 9, C. amycolatum ICIS 53, C. minutissimum ICIS 5 and C. xerosis ICIS 99.

Inhibitory effect of metabolites from Corynebacterium strains on human bacterial pathogens

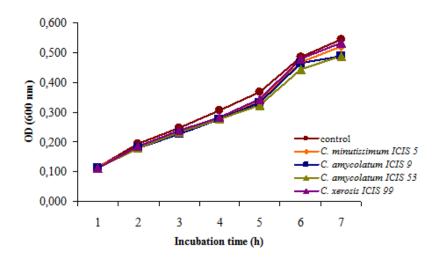
direct antagonism metabolites The of Corynebacterium strains against four human bacterial pathogens were studied using 96 wells polystyrene plate method. Figure 1-4, shows the optical density of K. pneumoniae, E. coli, P. aeruginosa and S. aureus grown in media containing the metabolites from different strains of Corynebacterium. The absorbance values demonstrate that all pathogenic strains were able to grow in media containing the metabolites of Corynebacterium. However, the pathogenic strains exhibited slower growth rate and reached lower final OD values of metabolites from Corynebacterium compared to the control.



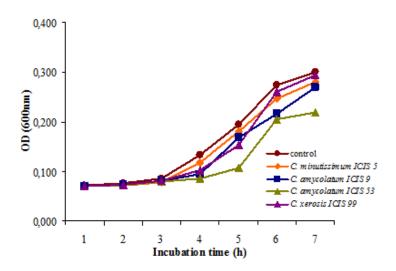
Graph 1
Growth of K. pneumoniae in cultural media containing the metabolites from Corynebacterium strains.



Graph 2
Growth of E. coli in cultural media containing the metabolites
from Corynebacterium strains.



Graph 3
Growth of *P. aeruginosa* in cultural media containing the metabolites from *Corynebacterium* strains.

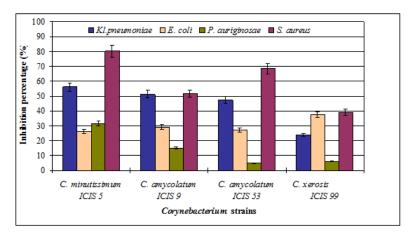


Graph 4
Growth of S. aureus in cultural media containing the metabolites from Corynebacterium strains.

Inhibitory effects of metabolites from Corynebacterium on biofilm formation of human bacterial pathogens

The results of inhibitory effects of metabolites from Corynebacterium strains on biofilm formation of human bacterial pathogens presented in Graph - 5. The metabolites from all four Corynebacterium strains inhibition of the biofilm formation all test bacterial pathogens, but the degree of inhibition of the biofilm formation varied among the Corynebacterium strains. results showed that all the Corynebacterium strains, except C. xerosis ICIS 99, exhibited strong inhibition of the biofilm formation of K. pneumoniae and S. aureus. Weak inhibition of the biofilm formation all the isolated Corynebacterium strains showed against P. aeruginosa. The metabolites

from *C.minutissimum ICIS* 5 inhibited biofilm formation of *K. pneumoniae* (56.1%), *E. coli* (26.3%), *P. aeruginosa* (31.7%) and *S. aureus* (80.3%). The metabolites from *C. amycolatum ICIS* 9 caused inhibition of biofilm formation *KI. pneumoniae* (51.3%), *E. coli* (29.2%), *P. aeruginosa* (15.2%) and *S. aureus* (51.7%). The metabolites from *C. amycolatum ICIS* 53 inhibited of the biofilm formation of *K. pneumoniae* (47.2%), *E. coli* (27.2%), *P. aeruginosa* (4.9%), and *S. aureus* (68.4%). Among the all *Corynebacterium strains*, the metabolites from *C. xerosis ICIS* 99 in less degree inhibited biofilm formation *K. pneumoniae* (23.7%) and *S. aureus* (39.1%). This strain inhibited biofilm formation of *E. coli* up to 37.5% and *P. aeruginosa* up to 6.3%

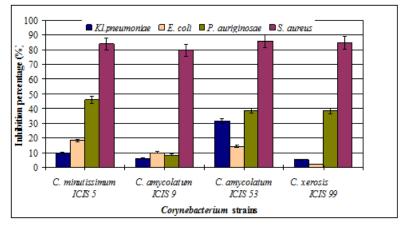


Graph 5
The influence of metabolites from Corynebacterium strains on biofilm formation of human bacterial pathogens. Data represent the mean and SD of three independent experiments each in triplicate format.

The influence of metabolites from Corynebacterium strains on pre-formed biofilms

The results of influence of metabolites from *Corynebacterium* strains on developed biofilms (24h grown) of human bacterial pathogens presented in Graph – 6. Treatment of preformed biofilms of *K. pneumoniae*, *E. coli*, *P. aeruginosa* and *S. aureus* in polystyrene microtiter plates with metabolites from all four *Corynebacterium* strains resulted in biofilm

disruption. To a greater extent this effect was observed under the influence metabolites from all four *Corynebacterium* strains on preformed biofilms *S. aureus* and *P. aeruginosa* (79.4 to 85.6% and 8.5 to 45.9%, respectively). To a lesser extent, metabolites from *Corynebacterium* strains influenced on preformed biofilms *K. Pneumoniae* and *E. coli* (5.1% to 31.6% and 2.4% to 18.5%, respectively).



Graph 6

The influence of metabolites from Corynebacterium strains on developed biofilms (24h grown) of human bacterial pathogens. Data represent the mean and SD of three independent experiments each in triplicate format.

DISCUSSIONS

The vaginal ecosystem is dynamic and contains microbiota, that are protective against invading pathogens, including those causing urinary tract infections and sexually transmitted infections. Lactobacillus strains are thought to play a major role in protecting in the microenvironment of the vagina and in inhibiting the overgrowth of potentially pathogenic organisms. ^{22,23,24} Using culture-independent techniques several investigators have demonstrated that a significant proportion (7-33%) of healthy women lack appreciable numbers of Lactobacillus species in the vagina 25,26 which may be replaced by other lactic acid producing bacteria such as Atopobium vaginae, Megasphaera, Leptotrichia species and Corynebacterium. 27 Although the structure of the communities may differ between populations, health can be maintained provided the function of these communities i.e. the production of lactic acid, continues. Consequently, the absence of lactobacilli or the presence of certain organisms such as G.vaginalis, of Peptostreptococcus, species Pseudomonas, Streptococcus and/or Corynebacterium does not constitute an abnormal state. 18,26 In this case, it is probably these microorganisms play the role in the protection of the vaginal biotope from pathogens. In present investigation, we study the antibacterial activity of metabolites from Corynebacterium spp. strains isolated from the reproductive tract of a healthy woman against four human pathogenic bacteria. The all Corynebacterium strains showed significant influence on growth, biofilm formation and on one-day developed biofilms K. pneumoniae, E. coli, P. aeruginosa and S. aureus. Decrease in growth all pathogenic strains in media containing the metabolites of Corynebacterium is probably due to the ability of strains of *Corynebacterium* to produce organic acids. Funke *et al.*, ²⁸ described, that the types of organic acids generated in the fermentation process depends on Corynebacterium species or strains, culture composition and growth conditions. The antimicrobial effect of organic acids lies in the reduction of pH, as well as the un-dissociated form of the molecules. It has been proposed that the low external pH causes acidification of the cell cytoplasm, while the undissociated acid, being lipophilic, can diffuse passively across the membrane. The un-dissociated acid acts by collapsing the electrochemical proton gradient, or by altering the cell membrane permeability, which results in disruption of substrate transport Besides, the antibacterial activity of metabolites of Corynebacterium may often be due to the production of bacteriocins or bacteriocin-like compounds. Nakano et al., 30 discovered in C.hydrocarboclastus the presence of Corynecin (an analog of chloramphenicol), characterized bactericidal activity against Gram positive and Gram negative bacteria (cocci and rods). Kwaszewska et al., determined the presence of a bacteriocin-like substance in 90% of the investigated Corynebacteria isolated from the skin of patients. The spectrum of its activity was covered by Gram positive bacteria, in particular S. aureus. Probably, thanks to the presence

of bacteriocins, the corynebacteria are able not only to suppress the increase in the biomass of the pathogenic bacteria, but also to affect biofilm formation. It is known that the biological activity of bacteriocins depends on the pH level.³² Probably, the corynebacteria isolated under normocenosis conditions influence the biofilm formation of pathogenic bacteria by synthesizing bacteriocins, while creating optimal conditions for their biological activity due to the acidification of the vaginal medium by the formation of organic acids. In addition, the ability of corvnebacteria to inhibit biofilm formation and destroy developed (24h grown) biofilms pneumoniae, E. coli, P. aeruginosa and S. aureus can also be associated with the production of surface-active substances (surfactants) of different nature. Zajic et al. ³³ found that *C. hydrocarboclastus* produces compounds having both surfactant and emulsifying properties. Thavasi et al. 34 demonstrated the emulsification capacities of the biosurfactant produced by C. kutscheri against hydrocarbons. Dalili et al. ¹⁷ described the lipopeptide coryxin isolated from C. xerosis NS5, isolated from the cubital fossa of a healthy person, which significantly suppressed the adhesive ability and biofilm formation S. aureus, Streptococcus mutans, E. coli and P. aeruginosa. The ability of Corynebacteria to influence growth, biofilm formation, and also to destroy biofilms of pathogenic microorganisms probably reveals one of the mechanisms of formation of normocenosis and determines the key role of Corynebacteria in protecting the vaginal biotope from infection in those women who do not have lactobacilli. In conclusion, the suggest a probiotic potential of Corynebacterium strains as an anti-bacterial agent in the vagina and in vivo studies, such as clinical trials designed to test their capacity to prevent and control urogenital tract infections in females.

CONCLUSION

In present study, we have reported the antibacterial activity of metabolites from *Corynebacterium spp.* strains isolated from the reproductive tract of a healthy woman against four human pathogenic bacteria. The all *Corynebacterium* strains showed significant influence on growth, biofilm formation and on developed biofilms (24h grown) *K. pneumoniae, E. coli, P. aeruginosa* and *S. aureus.* So, there is a need for further studies to isolate and characterize the bioactive compounds present in metabolites of *Corynebacterium* strains and these bioactive compounds can be used to develop effective drugs against these human pathogenic bacterial strains.

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CONFLICT OF INTEREST

Conflict of interest declared none.

REFERENCES

- Goodfellow M, Jones AL. Order V. Corynebacteriales ord. nov. In Goodfellow M, Kämpfer P, Busse H-J, Trujillo ME, Suzuki K-I, Ludwig W, Whitman WB (ed), Bergey's manual of systematic bacteriology, vol 5, 2nd ed. Springer Verlag, New York, NY; 2012. p. 232–243.
- 2. Sangal V, Hoskisson PA. Evolution, epidemiology and diversity of *Corynebacterium diphtheriae*: New perspectives on an old foe. Infect Genet Evol. 2016 Sep 43:364-70.
- 3. Bernard K. The Genus Corynebacterium and Other Medically Relevant Coryneform-Like Bacteria. J Clin Microbiol. 2012; 50(10):3152-58.
- Uehara Y, Nakama H, Agematsu K, Uchida M, Kawakami Y, Abdul Fattah AS, Maruchi N. Bacterial interference among nasal inhabitants: eradication of *Staphylococcus aureus* from nasal cavities by artificial implantation of *Corynebacterium* sp. J Hosp Infect. 2000 Feb 44(2):127-33.
- Bomar L, Brugger SD, Yost BH, Davies SS, Lemon KP. Corynebacterium accolens releases antipneumococcal free fatty acids from human nostril and skin surface triacylglycerols. MBio. 2016 Jan 5;7(1):e01725-15.
- Wysocki P, Kwaszewska AK, Szewczyk EM. Influence of substances produced by lipophilic Corynebacterium CDC G1 ZMF 3P13 on the microorganisms inhabiting human skin. Med Dosw Mikrobiol. 2011; 63(1):45-52.
- 7. Matthew M. Ramsey, Marcelo O. Freire, Rebecca A. Gabrilska, Kendra P. Rumbaugh, Katherine P. Lemon. *Staphylococcus aureus* shifts toward commensalism in response to *Corynebacterium* Species. Front Microbiol. 2016 Aug 17;7:1230.
- 8. Ma B, Forney LJ, Ravel J. Vaginal microbiome: rethinking health and disease. Annu Rev Microbiol. 2012; 66:371-89.
- Chaban B, Links MG, Jayaprakash TP, Wagner EC, Bourque DK, Lohn Z, Albert AY, van Schalkwyk J, Reid G, Hemmingsen SM, Hill JE, Money DM. Characterization of the vaginal microbiota of healthy Canadian women through the menstrual cycle. Microbiome. 2014 Jul 4;2(1):23.
- Pearce MM, Hilt EE, Rosenfeld AB, Zilliox MJ, Thomas-White K, Fok C, Kliethermes S, Schreckenberger PC, Brubaker L, Gai X, Wolfe AJ. The female urinary microbiome: a comparison of women with and without urgency urinary incontinence. MBio. 2014 Jul 8;5(4):e01283-14.
- Kokhreidze NA, Kravchenko ME. The formation of the vaginal microflora in the age aspect. J Children's Hospital. 2002; 3:45-47. (Article in Russian).
- Aagaard K, Riehle K, Ma J, Segata N, Mistretta TA, Coarfa C, Raza S, Rosenbaum S, Van den Veyver I, Milosavljevic A, Gevers D, Huttenhower C, Petrosino J, Versalovic J. A metagenomic approach to characterization of the vaginal microbiome signature in pregnancy. PLoS One. 2012; 7(6):e36466.

- Cherkasov SV, Gladysheva IV, Bukharin OV. Symbiotic interactions of corynebacteria and lactobacilli in realization of oxidative mechanisms of antagonism. Zh Mikrobiol Epidemiol Immunobiol. 2012 Nov-Dec (6):13-6.
- Prabakaran R and Kalimuthu K. Antibacterial activity of the whole plant of *Caralluma nilagiriana* kumari et subba rao – an endemic medical plant species. Int J Pharm Bio Sci. 2013; 4(1): (B) 42 – 48
- 15. O'Toole GA, Kolter R. Flagellar and twitching motility are necessary for *Pseudomonas aeruginosa* biofilm development. Mol Microbiol. 1998 Oct 30(2):295-304.
- Wojnicz D, Tichaczek-Goska D. Effect of subminimum inhibitory concentrations of ciprofloxacin, amikacin and colistin on biofilm formation and virulence factors of Escherichia coli planktonic and biofilm forms isolated from human urine. Braz J Microbiol. 2013 May 31;44(1):259-65.
- Dalili D, Amini M, Faramarzi MA, Fazeli MR, Khoshayand MR, Samadi N. Isolation and structural characterization of Coryxin, a novel cycliclipopeptide from *Corynebacterium xerosis* NS5 having emulsifying and anti-biofilm activity. Colloids Surf B Biointerfaces. 2015 Nov 1;135:425-32.
- Zhou X, Bent SJ, Schneider MG, Davis CC, Islam MR, Forney LJ. Characterization of vaginal microbial communities in adult healthy women using cultivation-independent. Microbiology. 2004 Aug 150(Pt 8):2565-73.
- 19. Pybus V, Onderdonk AB. Microbial interactions in the vaginal ecosystem, with emphasis on the pathogenesis of bacterial vaginosis. Microbes Infect. 1999 Apr 1(4):285-92.
- Cherpes TL, Meyn LA, Krohn MA, Lurie JG, Hillier SL. Association between acquisition of herpes simplex virus type 2 in women and bacterial vaginosis. Clin Infect Dis. 2003 Aug 1;37(3):319-25
- 21. Martin HL, Richardson BA, Nyange PM, Lavreys L, Hillier SL, Chohan B, Mandaliya K, Ndinya-Achola JO, Bwayo J, Kreiss J. Vaginal lactobacilli, microbial flora, and risk of human immunodeficiency virus type 1 and sexually transmitted disease acquisition. J Infect Dis. 1999 Dec 180(6):1863-8.
- 22. Gupta K, Stapleton AE, Hooton TM, Roberts PL, Fennell CL, Stamm WE. Inverse association of H₂O₂-producing lactobacilli and vaginal *Escherichia coli* colonization in women with recurrent urinary tract infections. J Infect Dis. 1998 Aug 178(2):446-50.
- 23. Klebanoff SJ, Hillier SL, Eschenbach DA, Waltersdorph AM. Control of the microbial flora of the vagina by H2O2-generating lactobacilli. J Infect Dis. 1991 Jul 164(1):94-100.
- Kaewsrichan J, Peeyananjarassri K, Kongprasertkit J. Selection and identification of anaerobic lactobacilli producing inhibitory compounds against vaginal pathogens. FEMS Immunol Med Microbiol. 2006 Oct 48(1):75-83.

- 25. Verhelst R, Verstraelen H, Claeys G, Verschraegen G, Delanghe J, Van Simaey L, De Ganck C, Temmerman M, Vaneechoutte M. Cloning of 16S rRNA genes amplified from normal and disturbed vaginal microflora suggests a strong association between *Atopobium vaginae*, *Gardnerella vaginalis* and bacterial vaginosis. BMC Microbiol. 2004 Apr 21;4:16.
- Hyman RW, Fukushima M, Diamond L, Kumm J, Giudice LC, Davis RW. Microbes on the human vaginal epithelium. Proc Natl Acad Sci USA. 2005 May 31;102(22):7952-7.
- Zhou X, Brown CJ, Abdo Z, Davis CC, Hansmann MA, Joyce P, Foster JA, Forney LJ. Differences in the composition of vaginal microbial communities found in healthy Caucasian and black women. ISME J. 2007 Jun 1(2):121-33.
- Funke G, von Graevenitz A, Clarridge J, III, Bernard KA. Clinical microbiology of coryneform bacteria. Clin Microbiol Rev. 1997 Jan 10(1):125-59
- Ammor S, Tauveron G, Dufour E. and Chevallier
 I. Antibacterial activity of lactic acid bacteria against spoilage and pathogenic bacteria isolated from the same meat small-scale facility: 1 –

- Screening and characterization of the antibacterial compounds. Food Control. 2006; 17:454–461.
- Nakano H, Tomita F, Yamaguchi K, Nagashima M, Suzuki T. Corynecin (chloramphenicol analogs) fermentation studies: selective production of Corynecin I by Corynebacterium hydrocarboclastus grown on acetate. Biotechnol Bioeng. 1977 Jul 19(7):1009-18.
- Kwaszewska A, Szewczyk EM. Production of antibacterial substances by resident corynebacteria isolated from human skin. Med Dosw Mikrobiol. 2007; 59:251–257.
- 32. Drosinos EH, Mataragas M, Nasis P, Galiotou M, Metaxopoulos J. Growth and bacteriocin production kinetics of *Leuconostoc mesenteroides* E131. J Appl Microbiol. 2005; 99(6):1314-23.
- 33. Zajic JE, Guignard H, Gerson DF. Emulsifying and surface active agents from *Corynebacterium hydrocarboclastus*. Biotechnol Bioeng. 1977 Sep 19(9):1285-301.
- 34. Thavasi R1, Jayalakshmi S, Balasubramanian T, Banat IM. Biosurfactant production by *Corynebacterium kutscheri* from waste motor lubricant oil and peanut oil cake. Lett Appl Microbiol. 2007 Dec 45(6):686-91.

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