



PHYTOCHEMICAL CONSTITUENTS AND THE DISTRIBUTION OF OXYCAROTENOIDS IN DEHYDRATED GREEN LEAFY VEGETABLES OF DIFFERENT SOURCES

SHERENA P A^{1*}, P T ANNAMALA² AND J K MUKKADAN³,

^{*1} *Department of Biochemistry, Little Flower Medical Research Center, Angamaly, Kerala*

² *Adjunct Faculty, Department of Biochemistry, LFMRC, Angamaly, Kerala*

³ *Research Director, Little Flower Medical Research Center, Angamaly, Kerala*

ABSTRACT

Plants possess many phytochemicals such as carotenoids, ascorbic acid, α -tocopherol, flavonoids, chlorophylls and polyphenols, etc. Green vegetables are packed with antioxidants, nutrients and fibers. These are essential for maintaining various biochemical processes including reducing levels of LDL-cholesterol, improving vision, helping to manage weight and boosting immunity. This study was undertaken to determine the total content of polyphenols, flavanoids, chlorophylls and xanthophylls in 15 different leafy vegetables from different regions by spectrophotometric method. The distribution of major oxycarotenoids such as lutein, zeaxanthin and β -cryptoxanthin was also studied using high performance liquid chromatography. The analytical data showed that the total polyphenol content ranged from 0.79-7.3 g/100g; total flavonoid content ranged from 1.1-23.7 g/100g; total chlorophyll content ranged from 0.47-4.7g/100g and the total xanthophyll content ranged from 0.17-1.65g/100g. In the oxycarotenoid distribution the results showed that lutein content of leafy vegetables ranged from 0.03-0.64 g/100g. The analysis also showed the presence of zeaxanthin and β -cryptoxanthin in some leaves like coriander curry leaf, fenugreek (methi) and mint leaves. Furthermore, we can conclude that the selected 15 different leafy vegetables are nutritionally and medicinally important and those of Indian sources are rich in xanthophylls whereas of Egyptian sources are rich in polyphenols and flavanoids

KEYWORDS: *Green leafy vegetables, polyphenols, flavonoids, chlorophylls, xanthophylls*



SHERENA P A

Department of Biochemistry, Little Flower Medical Research Center, Angamaly, Kerala

Received on : 17-04-2017

Revised and Accepted on : 26-05-2017

DOI: <http://dx.doi.org/10.22376/ijpbs.2017.8.3.b334-343>

INTRODUCTION

Green leafy vegetables are the cheapest of all the vegetables within the reach of poor man, being richest in their nutritional value¹. Green leafy vegetables are used since ancient periods as source of food as they contain many nutrients and minerals which are helpful in maintaining human health. Plant foods are sources of energy, micronutrients and nutrients essential to health, in addition to phytochemicals with further health benefits including glycemic control, immune-stimulation or antioxidant activity^{2,3}. Green leafy vegetables provide fiber, vitamins and minerals. Moreover, these have been identified as good sources of natural antioxidants like tocopherols, vitamin C and polyphenols which help in for maintaining good health and also protect against coronary heart diseases and cancer. Phytonutrients present in green leafy vegetables produce many common health benefits like protection from eye problems, oxidative stress, iron deficiency, etc^{4,5}. Carotenoids are important constituent of green leafy vegetables⁶. Besides chlorophylls and anthocyanins, carotenoids belong to the most important and most frequent pigments found in nature, the main sources being yellow, orange, and red fruits and vegetables as well as green leafy vegetables. Carotenoids, fat soluble, colored pigments serve a variety of functions in cellular biology. Carotenoids (β -carotene, lycopene and lutein) present in fruits and vegetables have antioxidant functions such as scavenging of singlet oxygen and other electronically excited molecules and also reduce the progression of many degenerative diseases^{7,8}. Antioxidants that neutralize free radicals are also secondary constituents or metabolites found naturally in the body and in plants such as fruits and vegetables. Plants contain an array of antioxidant compounds such as carotenoids, flavonoids, related polyphenols, cinnamic acid, benzoic acid, folic acid, ascorbic acid, tocopherols and tocotrienols which prevent oxidation of the susceptible substrate⁹. Common antioxidants include vitamin A, vitamin C, vitamin E and certain compounds called carotenoids (like lutein and β -carotene)¹⁰. Free radicals such as peroxide, hydroperoxide or lipidperoxyl are scavenged by polyphenols and flavonoids thereby inhibit the oxidative mechanisms¹¹. Polyphenols scavenge free radicals (R*) possessing an unpaired electron either by donation of hydrogen or electrons, resulting in comparatively stable phenoxyl (PhO*) radicals (neutral (PhO*) or cationic (PhO+*) molecules, respectively), which are stabilized by delocalization of unpaired electrons around the aromatic ring^{12,13a,13b}. Carotenoids are natural pigments and the pattern of conjugated double bonds in the polyene backbone of carotenoids determines their light absorbing properties and influences the antioxidant activity of carotenoids^{14,15}. The antioxidant action of carotenoids is their ability to quench singlet oxygen via physical or chemical quenching. According to the chemical composition, carotenoids are categorized as either carotenes or xanthophylls (oxycarotenoids); the carotene group, such as β -carotene, α -carotene and lycopene, composed only of carbon and

hydrogen atoms, xanthophylls such as zeaxanthin, lutein, α and β -cryptoxanthin, carry at least one oxygen atom¹⁶. Oxygenated carotenoids, including lutein, which have little or no pro-vitamin A activity, have been recognized to have antioxidant activities¹⁷. Singlet oxygen generated in biological systems is capable of damaging proteins, lipids and DNA. Lutein and zeaxanthin absorb visible light and play a role in singlet-singlet energy transfer and the quenching of singlet oxygen^{18,19}. This ability may contribute to the protection of light-exposed tissue, skin and eyes, from light-induced damage²⁰. The present study was conducted with the objective to quantify the phytochemical constituents and the distribution of oxycarotenoids in the dried green leafy vegetables which are commonly consumed in India, Egypt and Thailand.

MATERIALS AND METHODS

MATERIALS

Sample collection

Dehydrated green leafy vegetables of Curry leaves, Methi leaves, Coriander leaves, Broccoli leaves are collected from South East Asia (Farm to Home, Bangalore); Bay, Oregano, Marjoram, Basil, Pandan, Sage and Thyme from Egypt regions (Pharaohs Herbs, Export herbs and spices, Egypt Invoice No. 85/2015 & 4/2016); Lemongrass, Kaffir lime, and Parsley leaves are from Thailand (Chaichada Co Ltd., Bangkok; Invoice No. CCDHBDP17031-006062 & 006085) region.

Solvents

Methanol, Ethyl alcohol, Acetone, Hexane, Ethyl acetate, Toluene were purchased from Merck.

Reagents

Folin and Ciocalteu, Sodium Nitrite, Aluminium chloride, Potassium hydroxide, Sodium carbonate and Sodium sulphate were also purchased from Merck.

Standards

Gallic acid and Quercetin were purchased from Sigma. Carotenoid standards such as Lutein, Zeaxanthin, β -carotene and β -cryptoxanthin were received from Omni Active Health Technologies, Mumbai.

All solvents were checked for their purity by GC Analysis. High purity water was prepared with a Millipore water purification system.

METHODS

Sample preparation

The powdered leafy material of 0.5 g was refluxed twice with 5 ml methanol and 50 ml water for 1 hr at 80°C. Cooled and filtered quantitatively to a 100 ml volumetric flask and diluted with water. 5 ml of the aliquot was pipetted into 25 ml volumetric flask and diluted with water.

Total Polyphenol Content

The total polyphenols in dried leaves was determined using spectrophotometric method²¹. The color development was carried out by mixing 1 ml of the sample solution, 1 ml of Folin-Ciocalteu reagent, 15 ml water and 3 ml of saturated NaHCO₃

solution. Blank was concomitantly prepared, containing 1 ml 70% methanol, 1 ml Folin-Ciocalteu reagent, 15 ml water and 3 ml of saturated NaHCO_3 solution. The samples were heated in a thermostat at 45°C for 45 min. The absorbance was measured at 750 nm. The same procedure was repeated for the standard solution of gallic acid. Based on the measured absorbance, the polyphenols are calculated and expressed in terms of gallic acid equivalent.

Total Flavonoid Content

The total flavonoids in dried leaves was determined using spectrophotometric method²². The reaction mixture was prepared by mixing 1 ml of the sample solution, 0.3 ml of 5% sodium nitrite, 0.3 ml 10% Aluminium chloride, 2 ml 1M sodium hydroxide and 15 ml water. Blank was concomitantly prepared, containing 1 ml 70% methanol, 0.3 ml of 5% sodium nitrite, 0.3 ml 10% Aluminium chloride, 2 ml 1M sodium hydroxide and 15 ml water. The absorbance was determined using spectrophotometer at 510 nm. The same procedure was repeated for the standard solution of quercetin. Based on the measured absorbance, the flavonoids are calculated and was expressed in terms of quercetin equivalent.

Total Chlorophyll Content

The content of total chlorophyll in dried leaves was determined using spectrophotometric method²³. The powdered leafy material 3g was sonicated 4-5 times with 50 ml methanol each time for 20 minutes. Filtered into a 250 ml standard flask and made up to the mark using methanol. 5ml of the aliquot was further diluted to 50ml with acetone. The absorbance was noted at 410nm, 430nm, 642.5nm, 660nm and 663nm.

Total Xanthophyll Content

The content of total xanthophylls and their distributions in dried leaves was determined using spectrophotometric method and HPLC methods²³.

Extraction Of Oxycarotenoids from dried leaves

A representative sample of the grinded leafy sample 3 g was refluxed with 10 ml Hexane, 6 ml Ethanol, 6 ml Acetone, 7ml Toluene and 2 ml 40% methanolic KOH in a waterbath kept at $56^\circ\text{C} \pm 0.5^\circ\text{C}$ for 20 minutes. The

sample was cooled and 30 ml hexane was added and diluted to the mark with 10% sodium sulphate solution. Allowed to stand in the dark for 1 hour for the phase separation to take place. Pipetted 1 ml of aliquot from the upper phase in to 50 ml volumetric flask and diluted to the mark with the solvent hexane: acetone: methanol(8:1:1). The absorbance of the solution was measured at 474 nm for total xanthophylls and 5 ml of the aliquot was taken from the upper phase, filtered the solution using 0.22 micron Millipore filter, injected 20 micro liter of the solution into HPLC. Individual standards of carotenoids were also prepared by dissolving the respective standards in ethylacetate and made 50-100ppm solution.

HPLC Instrumentation

The analysis of the dried leaves and carotenoid standards were carried out by normal phase HPLC analysis. The Agilent Technology 1260 HPLC system equipped with DAD, gradient pump, a degasser, column thermostat and an autosampler. The separation was carried out in a Kromasil silica analytical column 250 x 4.6 mm id; particle size 5 μm with a Silica guard column used for normal phase analysis. The mobile phase consisted of hexane and ethyl acetate (75/25). All solvents were filtered through a syringe filter prior to the analysis. The flow rate was 2 ml /min for total run time 45 min., the detector was set 445 nm and Chemstation software was used for data processing and the injection volume was 20 μL .

RESULTS AND DISCUSSION

Table 1 depicts data on the analysis of phytochemical constituents such as polyphenols, flavonoids, chlorophylls and xanthophyll content measured by Spectrophotometry. Table 2 shows the distribution of oxy carotenoids like beta cryptoxanthin, lutein and zeaxanthin by normal phase chromatography. Table 3 represents the list of screened leafy vegetables with details of its family, scientific name and its medicinal properties. Figures 1, 2 and 3 show the absorption spectrum of carotenoids and chlorophylls; Overlaid chromatograms of individual standards such as lutein, zeaxanthin, β -cryptoxanthin and typical HPLC profile of dried methi leaf respectively

Table 1
Phytochemical constituents such as Polyphenols, Flavanoids, Chlorophyll and Xanthophyll content in the dehydrated green leafy vegetables

No:	sample name	Total Polyphenol Content (g/100g)	Total Flavanoid Content (g/100g)	Total Chlorophyll Content (g/100g)	Total Xanthophyll Content (g/100g)
1	Basil	7.28	23.72	0.4605	0.4447
2	bay leaves	2.88	6.83	0.6830	0.1767
3	Broccoli	1.06	2.20	3.8822	0.6884
4	coriander	0.79	2.64	4.7647	1.6545
5	curry leaf	4.25	6.42	0.9495	1.3267
6	kaffir lime	1.50	2.64	1.4170	0.5287
7	lemon grass	0.91	1.66	0.8894	0.1265
8	Marjoram	5.50	19.18	1.0836	0.1617
9	Methi	1.84	4.44	3.9406	1.4519
10	Mint	7.01	22.50	4.7737	0.2942
11	Oregano	6.40	11.02	0.4797	0.1488
12	pandan leaves	1.04	1.05	1.9484	0.5693
13	parsely leaves	1.04	0.92	1.6262	0.2176
14	Sage	6.60	11.63	0.9775	0.4360
15	Spinach	0.84	2.08	4.4609	1.2021
16	Thyme	4.99	21.95	1.3167	0.3953

Table 2
Distribution of Oxycarotenoids such as Lutein, Zeaxanthin and β -cryptoxanthin content in the dehydrated green leafy vegetables by HPLC

No:	sample name	Lutein Content (g/100g)	Zeaxanthin Content (g/100g)	β -Cryptoxanthin Content (g/100g)
1	Basil leaves	0.123305	0.01368	0
1	Bay leaves	0.081731	0	0
2	Broccoli leaves	0.280396	0	0
3	Coriander leaves	0.641668	0.044655	0.001145
4	Curry leaves	0.611147	0	0.001196
5	kaffir lime leaves	0.203332	0	0
6	lemon grass	0.037542	0	0
7	Marjoram leaves	0.080173	0.013459	0
8	Methi leaves	0.546312	0	0.002617
9	Mint leaves	0.081888	0.023284	0
10	Oregano leaves	0.065739	0.016384	0
11	pandan leaves	0.194324	0	0
12	parsely leaves	0.052948	0.019797	0
13	Sage leaves	0.201082	0	0
14	Spinach leaves	0.483692	0	0
15	Thyme leaves	0.169247	0.032754	0

Table3
List of dehydrated green leafy vegetables with their Local name, ScientificName, and Health benefits

Sl No.	Local name	Scientific Name	Major Health benefits*
1	Basil	<i>Ocimum basilicum</i>	Oral care, relief from respiratory disorders, treatment of fever, asthma, lung disorders, heart diseases and stress ,controlling blood glucose levels and cholesterol levels
2	Bay leaf	<i>Laurusnobilis</i>	Detoxify the body, slow the aging process, speed wound healing, protect the body from bacterial infections, manage diabetes, improve heart function, reduce inflammation, alleviate respiratory problems, optimize digestion, and prevent certain types of cancer.
3	Broccoli	<i>Brassica oleracea var. italic</i>	Anti-cancer, especially in the prevention and treatment of gastric cancer, breast cancer
4	Coriander	<i>Coriander sativum</i>	Skin inflammation, high cholesterol levels, diarrhea, mouth ulcers, anemia, indigestion, menstrual disorders, smallpox,conjunctivitis,skin disorders, and blood sugar disorders also benefiting eye care.
5	Curry leaves	<i>Murraya koenigii</i>	Anti-diabetic, antioxidant, antimicrobial, anti-inflammatory, anti-carcinogenic and hepato-protective
6	Kaffir lime	<i>Citrus hystrix</i>	Oral health, detoxify the blood, boost skin health, improve digestion, ward off insects, lower inflammation, aid the immune system, reduce stress, and improve the health of the hair relief from anxiety and stress
7	Lemon grass	<i>Cymbopogon</i>	Relief from stomach disorders, insomnia, respiratory disorders, fever, aches, infections, rheumatism and edema, helps in maintaining optimum cholesterol levels, cellular health, nervous system, healthy skin and immune system. Stopsdiarrhea
8	Marjoram	<i>Origanum majorana</i>	Relieving digestive problems, efficient sedative, stimulates lactation and perspiration, helps in flushing toxins from the body and anti-inflammatory
9	Methi leaves	<i>Trigonella foenum-graecum</i>	Aromatic,cooling,laxative, skin andmucousmembranes, relieve irritationof theskin, alleviate swelling andpain and diabetes

10	Mint leaves	<i>Mentha piperita</i>	Proper digestion and weight loss, relief from nausea, depression, fatigue and headache, treatment of asthma, memory loss, and skin care problems
11	Oregano leaves	<i>Origanum vulgare</i>	Respiratory tract disorders, gastrointestinal (GI) disorders, menstrual cramps, urinary tract disorders and rejuvenation
12	Pandan leaves	<i>Pandanus amaryllifolius</i>	Relieves gastrointestinal cramps and spasms, gives energy to new mothers, provides calming or soothing effects, lowers blood pressure, reduces fever and detoxifies the body
13	Parsely leaf	<i>Petroselinum crispum</i>	Controlling cancer, diabetes, and rheumatoid arthritis, along with helping prevent osteoporosis., relief from gastrointestinal issues and strengthen the immune system
14	Sage	<i>Salvia officinalis</i>	To improve brain function, lower inflammation, prevent chronic diseases, boost the strength of the immune system, regulate proper digestion, alleviate skin conditions, increase the health and strength of bones, prevent the onset of diabetes and lowers cholesterol
15	Spinach	<i>Spinacia oleracea</i>	Anti-oxidant,antiproliferative, anti-inflammatory,antihistaminic, CNSdepressant, protectionagainstgamma radiation and hepatoprotective effect
16	Thyme	<i>Thymus vulgaris</i>	Reduce respiratory issues, boost the strength of the immune system, protect against chronic diseases, stimulate blood flow, prevent fungal infections, improve heart health, and relieve stress fights sore throats

(*source'Useful Plants Of India";,"Encyclopedia of Medicinal Plants" and www.organicfacts.com)

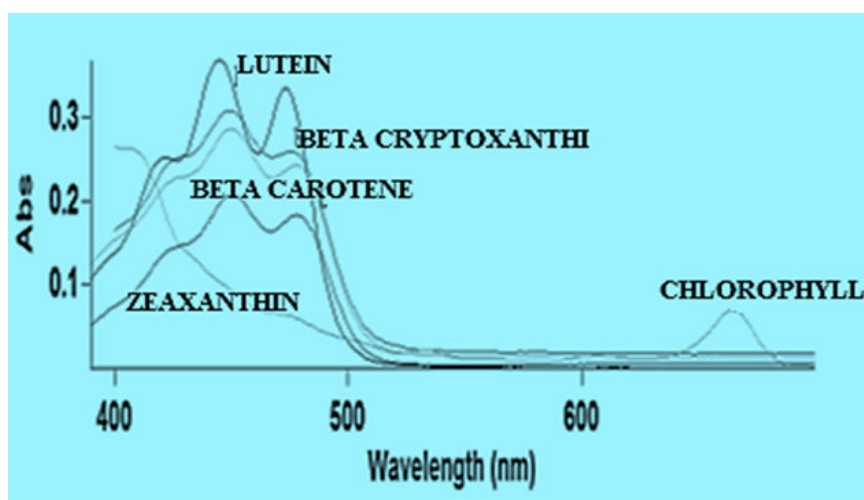


Figure 1
 Characteristic UV-Vis spectrum of Carotenoids (Lutein, zeaxanthin, beta carotene, beta cryptoxanthin) and chlorophyll

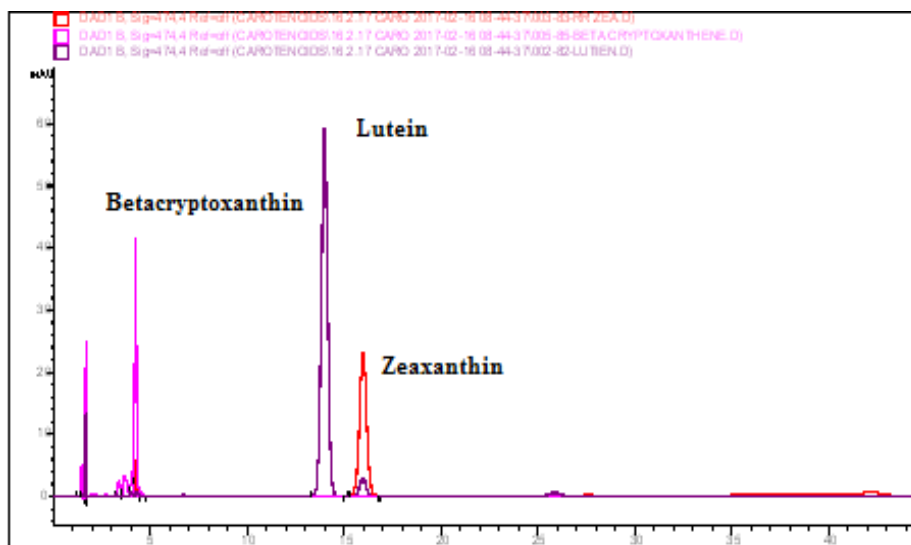


Figure 2
 HPLC Chromatogram of Mixed standards

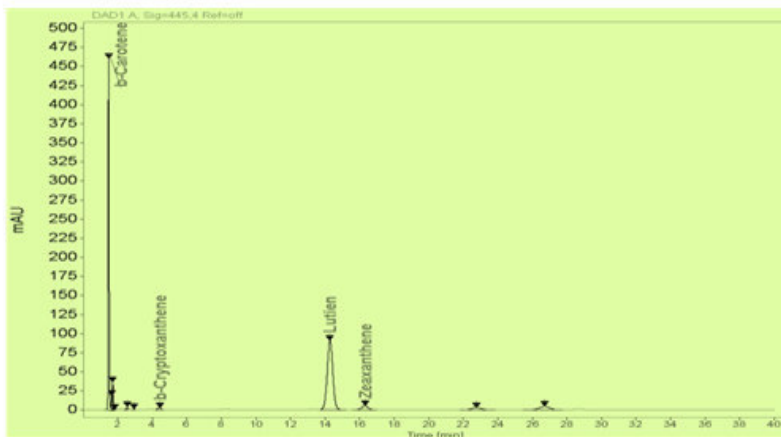
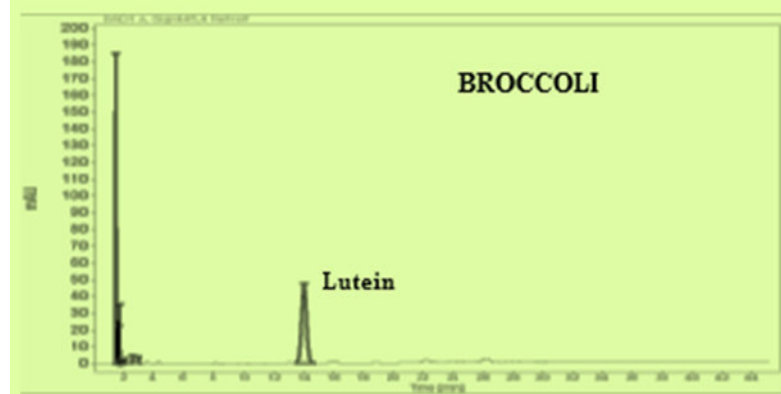
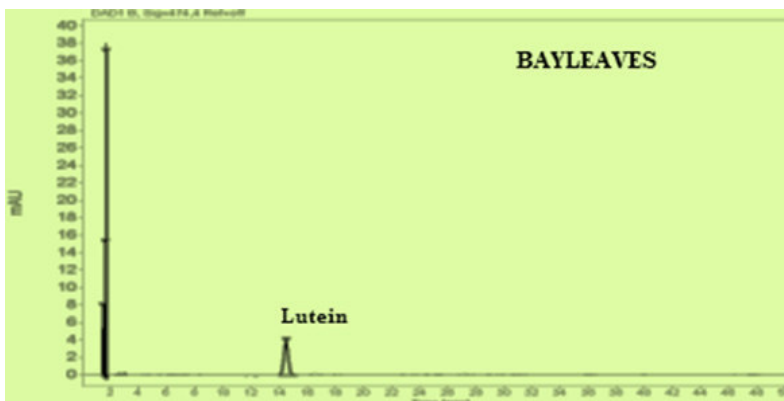
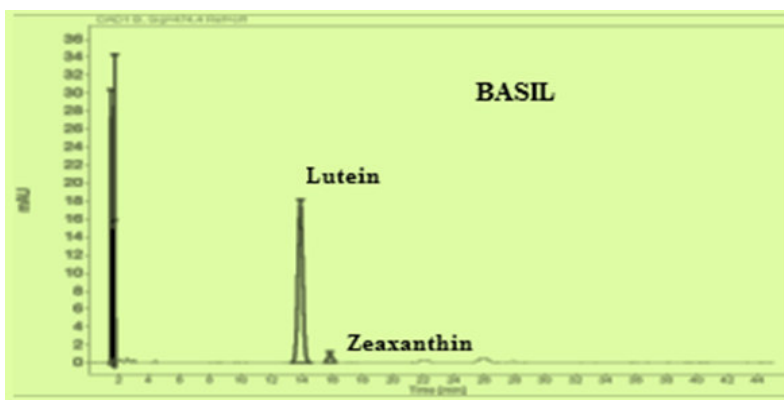
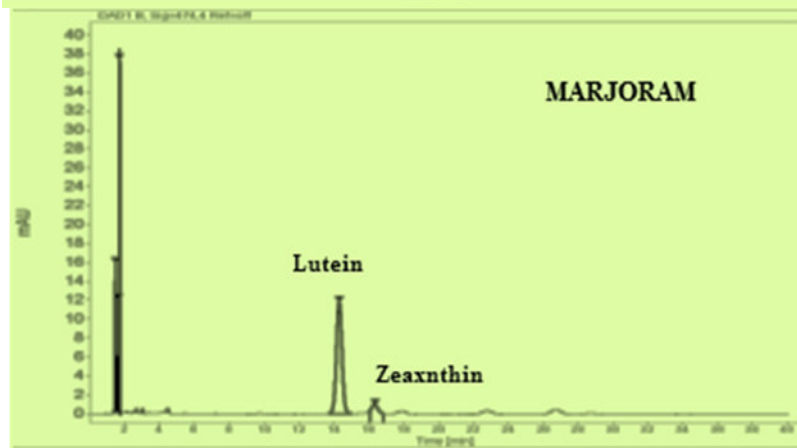
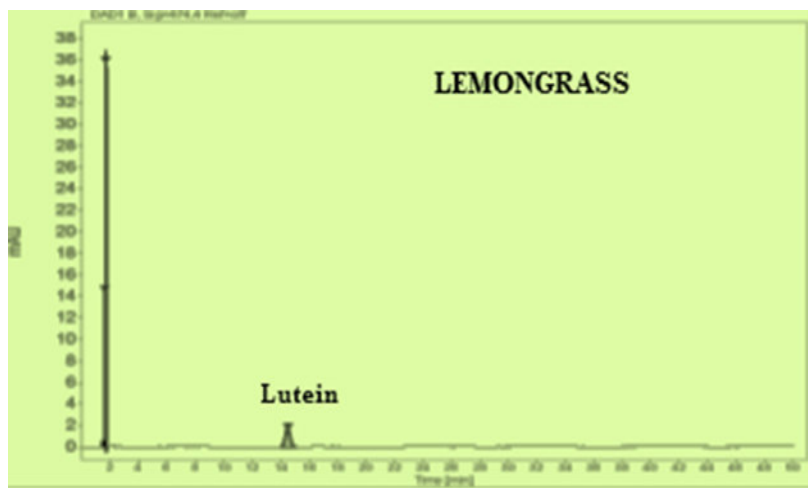
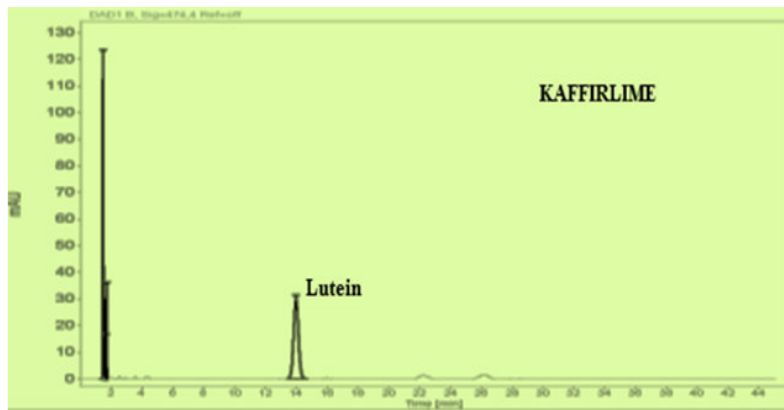
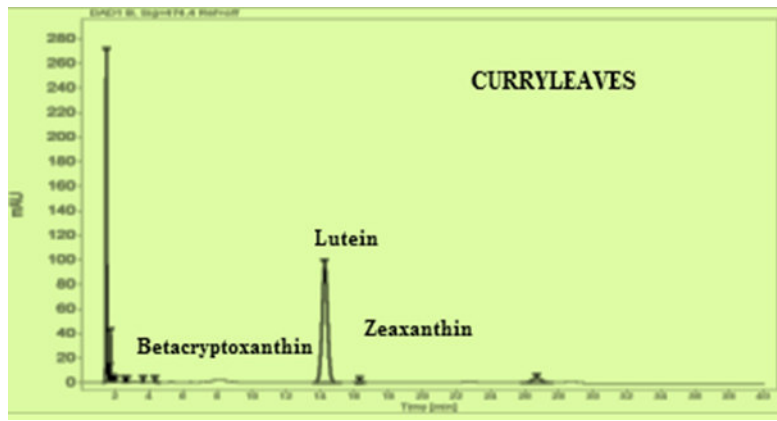
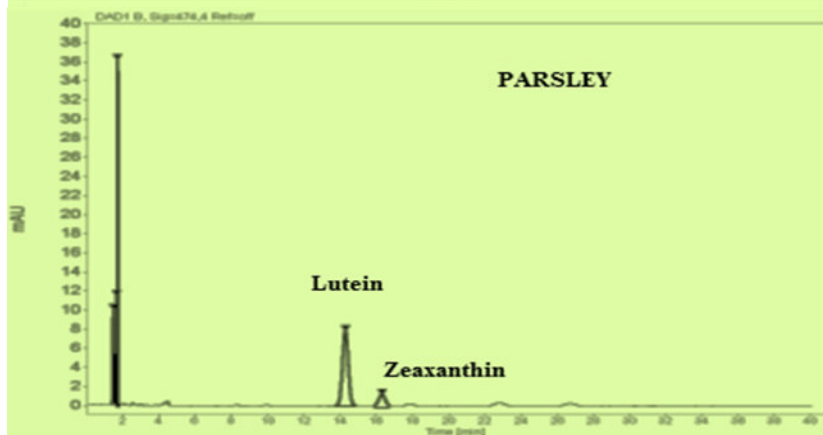
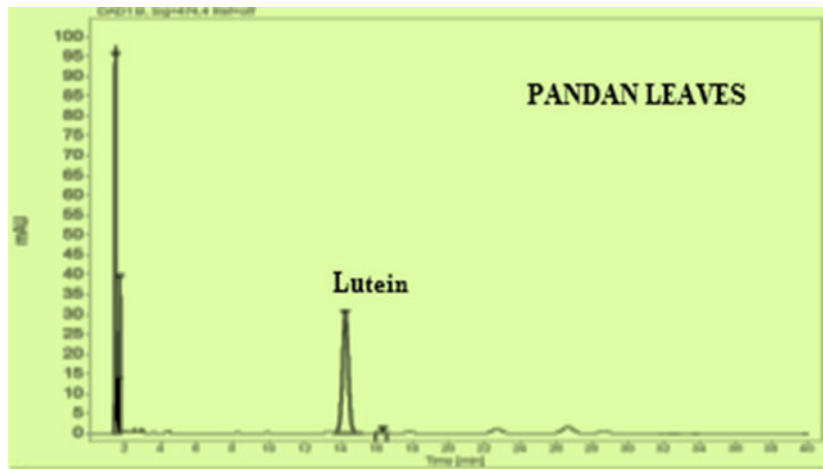
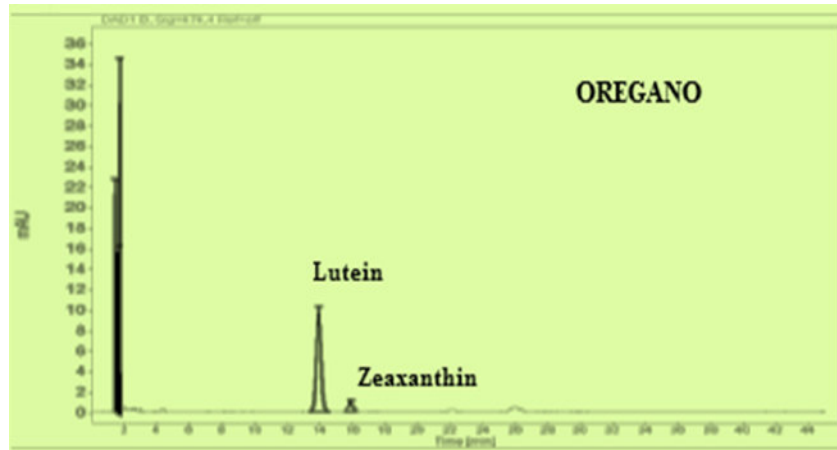


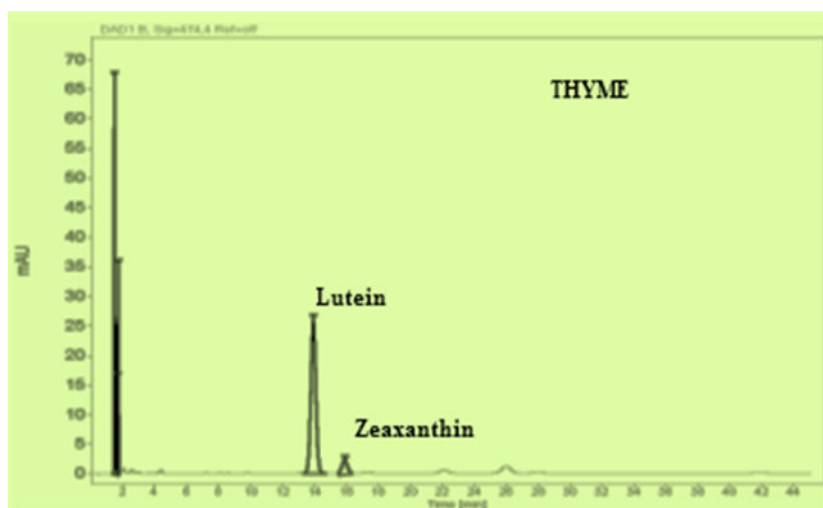
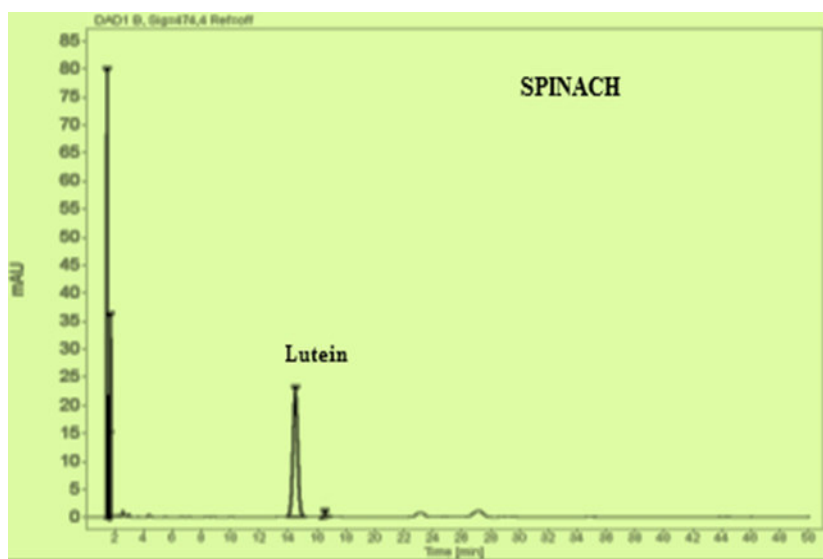
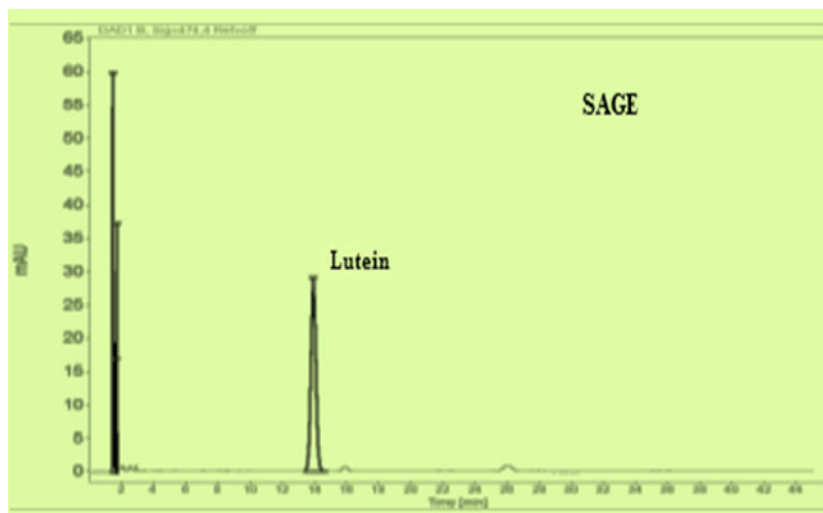
Figure 3
Typical HPLC profile of methileaves

APPENDIX:HPLC chromatogram of the selected leaves









The results of phytochemical constituents are presented in Table 2. From table 2 it is clear that the total polyphenols are in the range of 5-10% in basil, marjoram, mint, oregano and sage whereas in bayleaves, broccoli, curryleaves, kaffirlime, methi, pandan and parsley leaves are in between 1-5% and others such as coriander, lemongrass and spinach are showing less than 1%. The total flavonoids in dried leaves such as basil, marjoram, mint, oregano, sage and thyme are in the range of 10-24%; bayleaves and curryleaves are showing 5-10% and broccoli, coriander,

methi, kaffirlime, lemongrass, pandan, parsley and spinach are below 5%. Total chlorophyll contents are showing 3-5% in broccoli, coriander, methi, mint and spinach; 1-5% in kaffirlime, marjoram, parsley, pandan leaves and thyme; below 1% in basil, bayleaves, curryleaves, lemongrass, oregano and sage. Total xanthophyll contents are showing between 1-2% in coriander, curryleaves, methi and spinach; 0.5-1% in broccoli, kaffirlime and pandan leaves and others such as basil, bayleaves, lemongrass, marjoram, mint, oregano, parsley and thyme are below 0.5%. The data in

table 3 shows the content of carotenoids, in which lutein content in coriander, curryleaves and methi leaves are in the range of 0.5-1% while in broccoli, kaffirlime, spinach, sage are in the range of 0.2-0.5% and others like basil, bayleaves, lemongrass, marjoram, mint, oregano, parsley and thyme are 0.05-0.2%. Zeaxanthin content in basil, coriander, marjoram, mint, oregano, parsley and thyme are in the range of 0.01-0.08% and in others no zeaxanthin was observed. In most of the dried leaves betacyrptoxanthin was not showing any peak but in coriander, curry leaves and methi shows 0.00115, 0.00015, 0.0026% respectively.

CONCLUSION

The data on the phytochemical constituents and distribution of carotenoids shows that the selected leafy vegetables are nutritionally and medicinally important. Leafy vegetables of Indian sources such as

REFERENCES

- Kuhnlein, H. V. & ecveur, O. Dietary change and traditional food systems of indigenous peoples. Annual Review of Nutrition. 1996 July;16: 417-442).
- Ughade, S. N, Zodpey, S. P hanolkar, V. A. ,Risk factors for cataract: a case control study. Indian Journal of Ophthalmology. 1998 October;46(4): 221-227.2.
- Bélanger, J., Balakrishna, M., Latha, P., Katumalla, S., Johns, T. ,Contribution of selected wild and cultivated leafy vegetables from South India to lutein and β -carotene intake. Asia Pacific Journal of Clinical Nutrition. 2010 July ;19 (3): 417-424
- Gupta, S. Prakash, J., Nutritional and sensory quality of micronutrient-rich traditional products incorporated with green leafy vegetables. International Food Research Journal 2011 April;18(4): 667-675.
- Sommerburg, O., Keunen, J. E., Bird, A. C., Van Kuijk F. J. , Fruits and vegetables that are sources for lutein and zeaxanthin: The macular pigment in human eyes. British Journal of Ophthalmology. 1998 August; 82(8): 907-910.
- Devadas, R.P., Saroja S. Availability of Fe and β -carotene from Amaranthus to children. In P. Emmaus (Ed.), Proceedings of the second Amaranthus conference. 1980;15-21.
- Di Mascio P, Kaiser S Sies H . Lycopene as the most efficient biological carotenoid singlet oxygen quencher, Arch Biochem Biophys. 1989 November;274(2): 532.
- Sies H. Carotenoids, Antioxidants in disease, Mechanisms and Therapy, Academic press, New York ,USA, 1996
- Hollman P.C.H., Evidence for health benefits of plant phenols: Local or systemic effects? J. Sci. Food Agric. 2001 July;81(9):842-852.
- Hayek M. G., Massimino, S. P., Burr, J. R., Kearns, R. J. Dietary vitamin E improves immune function in cats. In G. A. Reinhart., & D. P. Carey (Eds.) Recent advances in canine and feline nutrition, 2000; Vol III p. 555-603
- Halliwell B, Auroama O.I. DNA and free radical mechanisms, CRC Press, Boca Raton, Florida, 579 pp: 1993
- Rice-Evans C.A, Miller N.J and Paganga G, Structure-antioxidant activity relationships of flavonoids and phenolic acids. J. Free Radic. Biol. Med., 1996 Feb;20(7):933-956.
- (a) Bouayed, J., H. Deuber, L. Hoffmann, T. Bohn, Bioaccessible and dialyzable polyphenols in selected apple varieties following in vitro digestion vs. their native patterns. Food Chem. 2011 April ;131(4): 1466-1472. (b) Bouayed J., L. Hoffmann T. Bohn, Antioxidative mechanisms of Whole-apple antioxidants employing different varieties from Luxembourg. J. Med. Food. 2011 December ; 14(12):1631-1637.
- Goodwin T.W. Carotenoids. In Encyclopedia of Plant Physiology, Bell E.A. and B.V. Charlwood (Eds.). Vol. 8, Springer-Verlag, Berlin, 1980 ; 257-281.
- Britton G., Liaaen-Jensen Sand Pfander H. Carotenoids, Biosynthesis and Metabolism, Vol. 3, Birkhauser Verlag, Basel, Switzerland, 1998 : 13-147
- Stahl W Sies H, Bioactivity and protective effects of natural carotenoids. Biochem. Biophys. Acta, 2005 May;740(2): 101-107.
- Krinsky, N.I. Actions of carotenoids in biological systems. Annual Review of Nutrition. 1993;13:561-587.
- Sadler M J, Strain J.J B. Caballero. Editors, Encyclopedia of Human Nutrition vol. 1, Academic Press, San Diego ,1999;1: 304-314.
- Di Mascio P, Devasagayam TP, Kaiser S, Sies H, Carotenoids, tocopherols and thiols as biological singlet molecular oxygen quenchers, Biochem Soc Trans. 1990 Dec;18(6):1054-6.
- Sies H, Stahl W, Non-nutritive bioactive constituents of plants: lycopene, lutein and

coriander, methi, curryleaves, spinach and broccoli are high in Chlorophyll, Xanthophylls and Lutein content whereas of Egyptian sources such as thyme, basil, oregano and marjoram are high in polyphenols and flavonoids.

ACKNOWLEDGEMENTS

The authors gratefully acknowledge the Kancor Ingredients Ltd for providing the facilities in the research division for carrying out this work. The authors thank Dr. T K Sunilkumar, Vice President R&D, Kancor Ingredients Ltd for his encouragement and support.

CONFLICT OF INTEREST

Conflict of interest declared none.

- zeaxanthin, Int J Vitam Nutr Res. 2003 Mar 73(2):95-100
21. Singleton V.I., Orthofer R., Lamuela-Raventos R.M., Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods Enzymol.*, 1999; 299: 152-178.
22. Quettier D.C., Gressier B., Vasseur J., Dine T., Brunet C., Luyckx M.C., Cayin J.C., Bailleul F., Trotin, F., Phenolic compounds and antioxidant activities of buckwheat (*Fagopyrum esculentum* Moench) hulls and flour. *J. Ethnopharmacol.*, 2000; 72: 35-42.
23. AOAC, Official methods of analysis of the Association of Official Analytical Chemists 17th Edition AOAC International, Gaithersburg MD. 2000.

Reviewers of this article



Dr. Wilma Delphine Silvia CR

Professor & Head, Dept. of Biochemistry,
Sapthagiri Institute of Medical Sciences and
Research Centre, Bangalore

Dr.Dinesha Ramadas Ph.D

Scientific Officer
Adichunchanagiri Institute for Molecular
Medicine
second floor,AIMS Building,AIMS campus
Nagamangala Taluk,Mandya



Asst.Prof.Dr. Sujata Bhattachary

Assistant Professor, School of Biological
and Environmental Sciences, Shoolini
University, Solan (HP)-173212, India



Prof.Dr.K.Suriaprabha

Asst. Editor , International Journal
of Pharma and Bio sciences.



Prof.P.Muthuprasanna

Managing Editor , International
Journal of Pharma and Bio sciences.

We sincerely thank the above reviewers for peer reviewing the manuscript