



PHENOLIC COMPOSITION AND ANTIOXIDANT ACTIVITY OF RAW SEEDS, GREEN SEEDS AND SPROUTS OF TEN FABA BEAN (*VICIA FABA*) CULTIVARS CONSUMED IN EGYPT

RAGAB EL-MERGAWI*^{1,2} AND HANAN A.A. TAIE³

¹Botany Department, National Research Centre, Cairo, Egypt, 12622.

²Plant Production and Protection Department, P. O. Box 6622, Buraydah, 51452, Saudi Arabia

³Plant Biochemistry Department, National Research Centre, Cairo, Egypt, 12622

ABSTRACT

Faba beans are widely consumed in Egypt as a vegetable by many ways. A study was conducted to determine the levels of phenolic compounds and antioxidant properties of raw seeds, green seeds and sprouts of ten faba bean cultivars commonly consumed in Egypt. Nubaria 2, Sakha 2 and Nubaria 1 cultivars showed higher amounts of phenolic compounds and exhibited the highest antioxidant activities (DPPH and FRAP). Four flavonoid constituents, myricetin, daidzein, apigenin and quercetin, were detected in tested faba bean seeds using HPLC with variable concentrations. For all tested cultivars, the mean values of total phenolics, tannins, total flavonoids, DPPH and FRAP in green seeds reached 342%, 485%, 267%, 439% and 136% relative to those found in raw seeds, respectively. The study concludes that the green faba bean seeds afforded an important source and good adequate level of antioxidants for daily inclusion in the human diet.

KEYWORDS: Phenolics, Antioxidant activity, Cultivars tannins, Green seeds, Sprouts, *Vicia faba*.



RAGAB EL-MERGAWI

Botany Department, National Research Centre, Cairo, Egypt, 12622.

*Corresponding author

INTRODUCTION

Faba bean (*Vicia faba* L.) is the most cultivated leguminous species in the world¹. Legumes are important component of traditional diets of several regions all over the world as they are low in fat, rich in protein, dietary fiber and a variety of micronutrients and phytochemicals². Egypt is one of the largest consumers of pulses in the world. Faba beans are a common staple food in the Egyptian diet, eaten by rich and poor alike. The most popular dishes of faba bean in Egypt are Medamis (stewed beans), Falafel (deep fried cotyledon paste with some vegetables and spices), Bissara (cotyledon paste poured onto plates) and Nabet soup (boiled germinated beans)³. Leguminous seeds, including faba bean belong to plant foods that are generally rich in phenolic compounds and possess high antioxidant capacity, which may be beneficial in the prevention of several health-related conditions like coronary and cardiovascular diseases, obesity, diabetes, inflammation and cancer^{4,5}. Phenolic compounds have antioxidant activity, which delays the oxidation of various "important for life" compounds by inhibiting the initiation or propagation of oxidising chain reactions. Natural antioxidants endogenous to food of plant origin can scavenge reactive oxygen and nitrogen species (RONS); evidence suggests that these may be of great importance in preventing the onset of oxidative diseases in the human body^{6,7}. Dry faba beans are widely known for their fiber, mineral and protein contents; however, its nutraceutical value is yet to gain as much attention in the prevention of chronic diseases⁸. Seeds of different faba beans cultivars are widely consumed in Egypt as a vegetable by many ways using dry seeds or green seeds or sprouts. There have been several studies of phenolic composition in dry faba bean seeds as this is the main means of consumption in the world^{9,10}. But, however, much less is known about the nutritional value of green seeds, which is considered as one way the faba beans are generally consumed in Egypt. Recently, differences in the phenolic composition and antioxidant activity between *V. faba* L. cultivars

were observed by Chaieb et al¹¹. and Baginsky et al¹². Sprouts are believed to be rich in health-promoting phytochemicals compare with their mature counterparts. Germination (sprouting) has been suggested as an inexpensive and effective way to improve the quality of legumes by increasing the levels of health-promoting antioxidants and decreasing of some antinutritional factors¹³. Cevallos-Casals and Cisneros-Zevallos¹⁴ examined 13 edible seeds for the levels of phenolic compounds and the antioxidant activity at different germination states (dormant, imbibed and 7d sprouts). They found that germinated edible seeds are an excellent source of dietary phenolic antioxidants. On the contrary, Al-Numair et al⁹ reported that, for two faba bean and three white bean cultivars, phenolic content decreased significantly throughout the sprouting periods up to 6 days. In order to demonstrate the health benefits of consuming faba beans with different ways, our objective was to determine the levels of different phenolic compounds, antioxidant properties and individualized flavonoid constituents by HPLC of raw seeds, green seeds and sprouts of ten faba bean cultivars commonly consumed in Egypt.

MATERIALS AND METHODS

(i) Chemicals and reagents

Flavonoid standards, including apigenin, isorhamnetin, kaempferol, luteolin, myricetin were purchased from Fluka (Bucks, Switzerland), quercetin, and neringenin were purchased from Sigma Chemical Co. (St. Louis, USA). Gallic acid, 1-1-diphenyl-2-picrylhydrazyl (DPPH), 6-hydroxy-2, 5, 7, 8-tetramethylchroman-2-carboxylic acid (Trolox) and 2, 4, 6-tripyridyl-s-triazine (TPTZ) were acquired from Sigma Chemical Co. (St. Louis, MO, U.S.A). Folin-Ciocalteu's reagent, methanol, hydrochloric acid, acetic acid, acetonitrile, sodium nitrite, aluminum chloride, sodium hydroxide, sodium acetate, ferric chloride, potassium dihydrogen phosphate,

butylated hydroxyanisole (BHA) and polyvinylpyrrolidone (PVPP) were purchased from Merck (Darmstadt, Germany). All reagents were of analytical grade or higher.

(i) Source and preparation of faba bean seeds

Raw faba bean seeds, which are the seeds of mature bean plants, of ten faba bean (*V. faba*) cultivars namely Giza 3, Giza 716, Giza 843, Nubaria 1, Nubaria 2, Sakha 1, Sakha 2, Sakha 3, Sakha 4, and Misr 1 were obtained from the Institute of Legumes in the Agriculture Research Centre, Giza, Egypt.

A germination test was conducted on ten faba bean cultivars. Seeds surface-sterilized with sodium hypochlorite (0.1%, w/v) for 2min, washed under running tap water for 5 min followed by distilled water and stored for further use. Five ml of distilled water were placed in a 9-cm plastic Petri dish lined with one Whatman No. 1 filter paper; ten seeds were placed on the paper. Petri dishes were kept in a growth chamber at 25 °C for three days (3d sprouts). Four replicates were maintained for each cultivar in a completely randomized manner. In all cultivars, 3d sprouts of each cultivar were stored in separate plastic bags at -20 °C prior to analyses. Seeds of ten tested cultivars were grown in green house of National Research Centre, Giza, Egypt. Mature green seeds were harvested for fresh consumption at the same physiological stage in three replicates of fifty plants each. Each replicate employed the same cultural practices, such as irrigation, fertilization, weeds and disease management, as are commonly used for *V. faba* cultivars in Egypt. Green pods with fully developed seeds were hand-harvested. In all cultivars, faba bean seeds of each replicate were stored in separate plastic bags at -20 °C prior to analyses.

(i) Seed extraction

Raw seeds, green seeds and sprouts, were collected, dried by a freeze-dryer (77500-00 M, Lab Conco Co, Mo, USA), finely grounded. Whole seed samples were initially crushed in a traditional stone mill and finally grounded with an analytic mill (IKA A11 basic; IKA Werke GmbH & Co. KG, Staufen, Germany). About

one gram of powdered sample was accurately weighed, and shaken with 250 ml of 80% ethanol for 12 hrs, using shaking incubator at room temperature. Solids were separated by centrifugation and filtration. Extracts were performed in triplicate for each individual cultivar, and subjected to determine antioxidant activity, total phenolics, tannins and total flavonoids.

(i) Determination of total phenolics tannins and flavonoids content

Total phenolic content was determined using Folin-Ciocalteu's reagent¹⁵ with some modifications. Seed extract (1 ml) was mixed with 1.5 ml of deionised water followed by 0.25 ml of Folin-Ciocalteu's reagent and allowed to react 6 min. Then 2.5 ml of sodium carbonate 7% was added and allowed to stand for 1 hr, then absorption at 765 nm was measured. Measurements were calibrated to a standard curve of prepared gallic acid solution, and the total phenolic was expressed as mg gallic acid equivalent (GAE)/ g of sample on a dry weight basis. Total tannin were determined in the extracts by a modification of the Folin-Ciocalteu's method using polyvinylpyrrolidone (PVPP) to separate tannin phenols from non-tannin phenols¹⁶. About 100 mg of PVPP was added to 1ml sample extract diluted with 1ml water and left 15 min at 4°C. After centrifugation, PVPP forms a precipitate with tannins, and the supernatant has only simple phenols. Simple phenols were determined using the Folin-Ciocalteu's reagent as previously mentioned. The difference between total and simple phenol values represents the tannin content, expressed as mg gallic acid equivalent (GAE)/ g of dried seeds. The flavonoid contents were measured using a colorimetric assay according to Zhishen et al¹⁷. A known volume (1 ml) of the extracts or standard solutions of quercetin was added to a 10 ml volumetric flask. Distilled water was added to make a volume of 5 ml. At zero time, 0.3 ml of 5% w/v sodium nitrite was added to the flask. After 5 min, 0.6 ml of 10% (w/v) AlCl₃ was added and, after 6 min, 2 ml of 1M NaOH were added to the mixture, followed by the addition of 2.1 ml distilled water. Absorbance

was read at 510 nm against the blank (water) and flavonoid content was expressed as mg quercetin equivalents (QE)/ g of dried seeds.

(i) Determination of antioxidant activity

The DPPH assay was done according to the method of Brand-Williams et al¹⁸ with some modifications. The stock solution was prepared by dissolving 24 mg 1, 1-diphenyl-2-picrylhydrazyl (DPPH) with 100 ml methanol and then stored at -20 °C until needed. The working solution was obtained by mixing 10 ml stock solution with 45 ml methanol to obtain an absorbance of 1.1±0.02 units at 515 nm using the spectrophotometer. Seed extracts (750 µl) were allowed to react with 1500 µl of the DPPH solution for 5 min in the dark. Then the absorbance was taken at 515 nm. The standard curve was linear between 25 and 800 µmol Trolox. Results are expressed in µmol Trolox/100g dry matter. Additional dilution was needed if the DPPH value measured was over the linear range of the standard curve. Ferric reducing antioxidant power assay (FRAP) was done according to Benzie and Strain¹⁹ with some modifications. The stock solutions included 300 mM acetate buffer (3.1 g C₂H₃O₂Na·3H₂O and 16 ml C₂H₄O₂), pH 3.6, 10 mM TPTZ (2, 4, 6-tripyridyl-s-triazine) solution in 40 mM HCl, and 20 mM FeCl₃·6H₂O solution. The fresh working solution was prepared by mixing 25 ml acetate buffer, 2.5 ml TPTZ solution, and 2.5 ml FeCl₃·6H₂O solution and then warmed at 37 °C before using. Seed extracts (150µl) were allowed to react with 2850 µl of the FRAP solution for 30 min in the dark condition. Readings of the colored product [ferrous tripyridyltriazine complex] were then taken at 593 nm. The standard curve was linear between 25 and 800 µM Trolox. Results are expressed in µmol Trolox/ 100g dry matter. Additional dilution was needed if the FRAP value measured was over the linear range of the standard curve.

(i) Analysis of flavonoid composition by HPLC

Flavonoid constituents were extracted according to McKeehen et al²⁰. Ground seeds were prepared as follows: 40 ml of 62.5%

aqueous methanol containing BHA (2g/ l) was added to 0.5 g dried sample material. To this, 10 ml of 6 N HCl was added to give a total volume of 50 ml. The extraction mixture was heated to 90 °C on a steam water bath and refluxed for 2 hrs, allowed to cool in the refrigerator, diluted to 100 ml with methanol and sonicated for 5 min to form the final extract. HPLC separation was carried out at a flow rate of 1ml/min and a temperature of 30 °C using a Shimadzu HPLC (Japan) with diode array detection and a supelcosil C₁₈ column (250 x 4.6 mm). The mobile phase was a linear gradient with O-phosphoric acid 0.25% (A)-acetonitrile (B) for 42 min starting with A:B (95:5) for 2 min, changing to A:B (90:10) for 5 min, A:B (85:15) for 3 min, A:B (80:20) for 13 min, A:B (70:30) for 5 min, A:B (50:50) for 4 min with equilibrating for 10 min. Detection was measured at 340 nm. Flavonoid standards, including apigenin, isorhamnetin, kaempferol, luteolin, myricetin, quercetin and neringenin. All the standards were dissolved in methanol to a concentration of 1 mg/10ml and were stored in darkness at -20 °C.

(i) Statistical analysis

Analyses were performed in triplicate and the data are presented as means. Analysis of variance and comparison of treatment means (LSD, 5% level) were performed using Stat graphics Plus Version 5.1.

RESULTS

1. Total phenolics

Total phenolics content in raw seeds, green seeds and 3d sprouts of ten faba bean cultivars are summarized in Table 1. Total phenolics in tested faba bean cultivars were significantly different and greatly influenced by seed form. Among test cultivars, phenolics in raw, green and sprouts are in the range 8.38 -14.01 mg GAE/g, 20.89- 46.58 mg GAE/g and 8.72-13.61 mg GAE/g, respectively. The greatest accumulation of phenolic compounds occurred in the green seeds. Since, the mean value of phenolics in green seeds for all tested cultivars was 35.6 mg GAE/g compared with 11.43 and

10.42 mg GAE/g for sprouts and raw seeds, respectively. Total phenolic content in the tested cultivars was significantly differed. In the raw seeds, the higher values were observed for Nubaria 2, followed by Misr 1 and Nubaria 1. But, in the green seeds, the phenolic contents of faba bean cultivars decreased as following Sakha 2 > Nubaria 2> Sakha 3> Sakha 4. Whereas, in the sprouts, Nubaria 1 and Nubaria 2 exhibited the high levels of total phenolics of all cultivars studied.

2. Tannins and flavonoids

As shown in Table 1, levels of tannins and total flavonoids presented a high variability according to seed cultivars and seed forms. The results indicated that the greatest accumulation of

tannins and flavonoids occurred in green seeds as compare with raw seeds or sprouts. Tannins in green seeds ranged from 9.5 to 20.98 mg GAE/g, in raw seeds ranged from 2.07 to 5.06 mg GAE/g and in sprouts ranged from 3.36 to 6.67 mg GAE/g. Whereas, total flavonoids in green seeds ranged from 4.27 to 13.20 mg QE/g, in raw seeds ranged from 2.73 to 4.73 mg QE/g and in sprouts ranged from 3.13 to 5.47 mg QE/g. Significant differences in the contents of tannins and flavonoids between tested cultivars were detected. In raw seeds and green seeds, Nubaria 2 and Sakha 2 cultivars accumulated high amounts of tannins and flavonoids. Whereas, in sprouts, the greatest accumulation of tannins and flavonoids occurred in Nubaria 1 and Nubaria 2 cultivars.

Table 1
Phenolic compounds in raw, green and sprouts of different faba bean cultivars.

Cultivars	Total phenolics (mg GAE/g DW)			Tannins (mg GAE/g DW)			Total flavonoids (mg QE/g DW)		
	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout
Giza 3	8.38 ^{dB}	36.20 ^{bA}	8.72 ^{EB}	2.56 ^{abB}	17.04 ^{abA}	3.36 ^{dB}	2.80 ^{CC}	9.07 ^{CA}	4.93 ^{abB}
Giza 716	9.31 ^{cdC}	20.89 ^{CA}	11.26 ^{bcB}	2.37 ^{abC}	9.50 ^{CA}	5.19 ^{bb}	3.47 ^{bcB}	4.27 ^{EA}	3.13 ^{dB}
Giza 843	9.64 ^{cdB}	33.51 ^{bA}	9.85 ^{dB}	2.55 ^{abB}	13.24 ^{bcA}	4.30 ^{CB}	2.73 ^{cC}	8.53 ^{CA}	3.60 ^{bcdB}
Nubaria 1	10.95 ^{bcC}	33.15 ^{bA}	13.61 ^{aB}	4.33 ^{abB}	17.28 ^{abA}	6.46 ^{ab}	3.87 ^{abcC}	7.87 ^{CA}	5.27 ^{aB}
Nubaria 2	14.01 ^{abB}	44.65 ^{aA}	13.33 ^{aB}	5.06 ^{ab}	20.25 ^{aA}	6.67 ^{ab}	4.73 ^{aB}	12.80 ^{abA}	5.47 ^{aB}
Sakha 1	9.64 ^{cdC}	23.33 ^{CA}	11.73 ^{bcB}	3.79 ^{abC}	13.21 ^{bcA}	6.03 ^{ab}	3.20 ^{CB}	5.87 ^{DA}	3.47 ^{bcdB}
Sakha 2	10.93 ^{bcB}	46.58 ^{aA}	11.35 ^{bcB}	4.02 ^{abB}	20.98 ^{aA}	6.26 ^{ab}	4.40 ^{abB}	13.20 ^{aA}	3.47 ^{bcdB}
Sakha 3	9.49 ^{cdB}	42.88 ^{aA}	10.33 ^{cdB}	3.30 ^{abB}	19.30 ^{abA}	4.39 ^{CB}	2.80 ^{cC}	12.40 ^{abA}	4.73 ^{abB}
Sakha 4	9.81 ^{cdC}	41.21 ^{aA}	12.24 ^{bB}	3.52 ^{abC}	19.59 ^{aA}	6.4 ^{ab}	3.47 ^{bcB}	11.07 ^{bA}	3.47 ^{bcdB}
Misr 1	12.03 ^{bb}	33.61 ^{bA}	11.84 ^{bcB}	2.07 ^{bC}	12.58 ^{bcA}	6.30 ^{ab}	3.53 ^{bcB}	8.53 ^{CA}	3.33 ^{cdB}

GAE: gallic acid equivalent; QE: Quercetin equivalent. The same small letters in a column are not significantly different ($p < 0.05$). The same capital letters in a row are not significantly different ($p < 0.05$).

Any symbol should be written in small

3. Antioxidant activity

The antioxidant activity of raw, green and sprouts for ten faba bean cultivars were assayed using DPPH and FRAP (Table 2). Significant variations in antioxidant activities were observed between cultivars and between seed forms. The DPPH values showed a more changeable behavior than FRAP values. Among tested cultivars, DPPH and FRAP values were in the range 2196- 5604 and 2333- 3391 $\mu\text{mol Trolox}/100\text{ g}$ for raw seeds, 5400 - 26880 and 3539 - 3852 $\mu\text{mol Trolox}/100\text{ g}$, for green seeds as well as 2520-5448 and 2639-3377 $\mu\text{mol Trolox}/100\text{ g}$ for sprouts, respectively. Green seeds exhibited the highest DPPH and FRAP

values (mean values 14724 and 3792 $\mu\text{mol Trolox}/100\text{ g}$, respectively) followed by sprouts (mean values 4096 and 2947 $\mu\text{mol Trolox}/100\text{ g}$, respectively) and finally raw seeds (mean values 3358 and 2789 $\mu\text{mol Trolox}/100\text{ g}$, respectively). Between raw seeds, the highest values of DPPH and FRAP occurred by Nubaria 2 cultivar. This cultivar had 155 % more DPPH activity than the Giza 3, the cultivar with the lowest value of DPPH activity and had 45 % more FRAP activity than the Giza 843, the cultivar with the lowest value of FRAP activity. In the green seeds, Sakha 2 showed the highest DPPH antioxidant activity, while, in sprouts, Nubaria 1 and Nubaria 2 cultivars had

the highest FRAP and DPPH activities of all cultivars studied.

Table 2
Antioxidant activity ($\mu\text{mol Trolox}/100 \text{ g DW}$) of raw, green and sprouts of different faba bean cultivars.

Cultivars	DPPH			FRAP		
	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout
Giza 3	2196 ^{cb}	14880 ^{bca}	2520 ^{db}	2509 ^{db}	3834 ^{ba}	2747 ^{cb}
Giza 716	2256 ^{cc}	6240 ^{ca}	3720 ^{cb}	2671 ^{db}	3539 ^{ca}	2891 ^{bcb}
Giza 843	2256 ^{cc}	7080 ^{ca}	3720 ^{cb}	2333 ^{bc}	3802 ^{ba}	2743 ^{cb}
Nubaria 1	3792 ^{bc}	12840 ^{bca}	5232 ^{ab}	2923 ^{bc}	3809 ^{ba}	3377 ^{ab}
Nubaria 2	5604 ^{ab}	21120 ^{aba}	5448 ^{ab}	3391 ^{ab}	3852 ^{ba}	3344 ^{ab}
Sakha 1	3336 ^{bcb}	5400 ^{ca}	4260 ^{bcb}	2711 ^{db}	3704 ^{ba}	2819 ^{bcb}
Sakha 2	3624 ^{db}	26880 ^{ba}	3600 ^{cb}	2866 ^{db}	3845 ^{ba}	2639 ^{cb}
Sakha 3	3300 ^{bcb}	17160 ^{abca}	4140 ^{bcb}	2768 ^{db}	3852 ^{ba}	2902 ^{bcb}
Sakha 4	3036 ^{bcb}	21000 ^{aba}	3888 ^{cb}	2765 ^{bc}	3852 ^{ba}	3186 ^{ab}
Misr 1	4176 ^{db}	14640 ^{bca}	4428 ^{abcb}	2956 ^{db}	3830 ^{ba}	2819 ^{bcb}

The same small letters in a column are not significantly different ($p < 0.05$).
The same capital letters in a row are not significantly different ($p < 0.05$).

4. Flavonoid constituents

Four flavonoids constituents, myricetin, daidzein, apigenin and quercetine, were detected in faba bean seeds (Table 3). Myricetin and daidzein found to be the main flavonoid components in raw seeds, green seeds and sprouts. Myricetin varied from

Table 3
Flavonoid constituents ($\mu\text{g/g DW}$) of raw, green and sprouts of different cultivars.

Cultivars	Myricetin			Daidzein			Apigenin			Quercetine		
	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout	Raw seed	Green seed	Sprout
Giza 3	5.42 \pm 0.41	3.86 \pm 0.32	7.83 \pm 0.44	1.22 \pm 0.08	1.16 \pm 0.03	7.39 \pm 0.43	0.85 \pm 0.01	0.54 \pm 0.01	1.50 \pm 0.01	0.56 \pm 0.01	0.61 \pm 0.01	4.15 \pm 0.12
Giza 716	4.58 \pm 0.21	3.38 \pm 0.23	7.60 \pm 0.43	2.67 \pm 0.04	1.44 \pm 0.02	7.23 \pm 0.32	1.99 \pm 0.01	ND	2.42 \pm 0.03	1.35 \pm 0.01	0.52 \pm 0.02	3.61 \pm 0.02
Giza 843	4.32 \pm 0.11	3.39 \pm 0.08	12.6 \pm 0.60	2.29 \pm 0.02	1.84 \pm 0.02	7.69 \pm 0.04	0.71 \pm 0.01	ND	2.12 \pm 0.02	2.36 \pm 0.04	ND	6.44 \pm 0.19
Nubaria 1	5.08 \pm 0.15	3.59 \pm 0.06	8.70 \pm 0.32	5.08 \pm 0.18	3.59 \pm 0.09	9.28 \pm 0.72	1.63 \pm 0.03	1.07 \pm 0.01	2.96 \pm 0.02	0.70 \pm 0.01	1.28 \pm 0.02	1.94 \pm 0.01
Nubaria 2	4.58 \pm 0.15	4.08 \pm 0.09	12.78 \pm 0.40	1.18 \pm 0.08	2.48 \pm 0.07	8.88 \pm 0.56	1.81 \pm 0.01	1.16 \pm 0.02	2.84 \pm 0.04	1.04 \pm 0.02	0.83 \pm 0.02	5.39 \pm 0.18
Sakha 1	4.14 \pm 0.08	3.82 \pm 0.09	6.12 \pm 0.29	1.81 \pm 0.05	1.45 \pm 0.03	4.95 \pm 0.16	ND	0.53 \pm 0.01	ND	0.84 \pm 0.02	0.72 \pm 0.02	7.24 \pm 0.81
Sakha 2	4.34 \pm 0.11	4.36 \pm 0.98	14.22 \pm 0.73	2.10 \pm 0.03	2.43 \pm 0.01	9.75 \pm 0.65	0.66 \pm 0.00	ND	3.08 \pm 0.05	0.81 \pm 0.01	0.68 \pm 0.01	4.31 \pm 0.31
Sakha 3	3.65 \pm 0.06	2.61 \pm 0.04	7.18 \pm 0.49	1.13 \pm 0.01	1.78 \pm 0.02	5.27 \pm 0.60	0.40 \pm 0.01	ND	1.55 \pm 0.03	0.56 \pm 0.01	0.86 \pm 0.01	5.97 \pm 0.22
Sakha 4	2.84 \pm 0.03	3.52 \pm 0.04	6.37 \pm 0.20	1.84 \pm 0.02	1.38 \pm 0.01	7.16 \pm 0.54	0.80 \pm 0.01	0.55 \pm 0.01	1.32 \pm 0.02	1.05 \pm 0.03	0.99 \pm 0.02	2.37 \pm 0.18
Misr 1	3.81 \pm 0.23	4.27 \pm 0.11	8.11 \pm 0.33	1.19 \pm 0.01	2.88 \pm 0.01	7.46 \pm 0.34	0.29 \pm 0.00	0.68 \pm 0.02	1.08 \pm 0.01	2.78 \pm 0.04	3.15 \pm 0.40	11.10 \pm 0.92

Values are means of three samples; ND: not identified.

2.84 to 5.42 ug/g in raw seeds, from 2.61 to 4.27 ug/g in green seeds and from 6.12 to 12.78 ug/g in sprouts. While, daidzein ranged from 2.1 to 5.08 ug/g in raw seeds, and from 11.6 to 3.59 ug/g in green seeds. Apigenin and quercetin appeared in small quantities in raw seeds or green seeds. For the same cultivar, the highest values for myricetin, daidzein, apigenin and quercetin were observed for the sprouts as compared with green seeds or raw seeds. Among studied cultivars, concentrations of flavonoid constituents showed considerable variations. Nubaria 1, Sakha 2 and Nubaria 2 seeds tended to contain higher concentrations of myricetin, daidzein and apigenin than the other tested cultivars. Whereas, Misr 1 recorded the highest concentration of quercetin constituent

DISCUSSION

Significant differences between faba bean cultivars were equally detected for all tested phenolic compounds and antioxidant activity, among the same seed form. Variations in phenolic content and antioxidant activity between faba beans cultivars were previously observed by Chaieb et al¹¹ and Ismael et al¹⁰. Our results about the levels of total phenolic and tannin contents are comparable with those obtained by Baginsky et al¹². They found that, among ten *V. faba* L. cultivars, total phenolics ranged from 817 to 1337 mg gallic acid equivalent per kilogram and condensed tannin content ranged from 309 to 958 mg (+)-catechin equivalent per kilogram. The differences in the concentration of phenolic compounds and antioxidant activity, among tested cultivars are likely to primarily reflect genotypic variation²¹. For all tested seeds, Nubaria 2, Sakha 2 and Nubaria 1 cultivars tended to contain higher amounts of tested phenolic compounds and better antioxidant activity. The large amount of variation observed among the ten cultivars is promising for future efforts to develop and improve strains of faba beans in Egypt for maximal phenolic content and antioxidant activity. A few studies have been performed on the antioxidant activity of faba beans¹¹. The

results of this study indicated that the seeds extract of faba bean was found to display a good antioxidant effect. The superiority of DPPH and FRAP antioxidant activities of faba bean in comparison with other legumes such as peas, lentils and soybean were observed by Lin & lai²¹ Lopez-Amoros et al²², and Sreeramulu et al²³. Also, Chaieb et al¹¹ reported that faba bean extracts had a higher capacity of scavenging the DPPH radical than synthetic antioxidants (BHT or TBHQ) and pure phenolic constituents as quercetin, catechin, rutin and gallic acid.

Presence of myricetin, apigenin and quercetin in faba bean seeds is consistent with others studies that have reported that in faba beans, the flavonols are mainly myricetin, kaempferol and quercetin, both free aglycones and glycosides^{24,25}. Also, Nubaria 1, Sakha 2 and Nubaria 2 seeds tended to contain higher concentrations of myricetin, daidzein and apigenin than the other tested cultivars. The observed differences in the concentration of flavonoid compounds among cultivars are likely to primarily reflect genotype variations¹². From our results, it can be observed that sprouts tended to accumulate more phenolics, tannins and flavonoids content and antioxidant activity than raw seeds. An increase of total phenols and antioxidant activity after germination was reported by Cevallos-Casals & Cisneros-Zevallos¹⁴ for selected seeds included mungbean, alfalfa, fava, fenugreek, mustard, wheat, broccoli, sunflower, soybean, radish, kale, lentil and onion. Duenas et al²⁶ reported that germination caused significant changes in the phenolic composition (increasing) due mainly to endogenous enzymes' activation and the complex biochemical metabolism of seeds during this process. The obtained results indicated that, for all faba bean cultivars, greatest accumulation of all tested phenolic compounds and antioxidant activity occurred in green seeds as compare with either raw seeds or sprouts. The mean value of green seeds showed 242%, 385%, 167%, 439% and 360% increases in total phenolics, tannins, total flavonoids, DPPH and FRAP relative to these found in raw seeds, respectively. In line to these results, Toma's-Barbera'n et al²⁵ reported that

the variations in phenolic constituents in faba bean seeds depended on the stage of maturity. There is a lack of information about phenolic composition and antioxidant activity of green seeds, which is considered as one way the faba beans are generally consumed in Egypt. Our results revealed that the green seeds are a good source of phenolics, the bioactive compounds with health promoting antioxidants.

CONCLUSION

This research is a contribution to the determine of the phenolic contents, antioxidant activity and individualized flavonoid constituents by HPLC

REFERENCES

1. Nadal S, Suso M J and Moreno M T, Management of *Vicia faba* L. genetic resources changes associated to the selfing process in the major, equina and minor groups, *Gentic Resources and Crop Evolution*, 50:183–192, (2003).
2. Kanatt SR, Chander R and Sharma A, Antioxidant potential of mint (*Mentha spicata* L.) in radiation processed lamb meat, *Food Chem*, 100: 451–458, (2007).
3. Jambunathan R, Blain HL, Dhindsa KH, Hussein LA, Kogure K, Li-Juan L, and Youssef MM, Diversifying use of cool season food legumes through processing. In: F.J. Muehlbauer and W.J. Kaiser (eds.), *Expanding the Production and Use of Cool Season Food Legumes*. Kluwer Academic Publishers, Dordrecht. The Netherlands, 1994, pp 98-112.
4. Losso JN, Targeting excessive angiogenesis with functional foods and nutraceuticals, *Trends Food Sci Tech*, 14:455–468, (2003).
5. Caccialupi P, Ceci LR, Siciliano RA, Pignone D, Clemente A and Sonnante G, Bowman-Birk inhibitors in lentil: heterologous expression, functional characterization and anti-proliferative properties in human colon cancer cells, *Food Chem*, 120:1058–1066, (2010).
6. Halliwell B, Gutteridge J M C and Cross C E, Free radicals, antioxidants, and human disease: where are we now?, *J Lab Clin Medicine*, 119:598–620, (1992).
7. Willett WC, Diet and health: What should we eat?, *Science* 264:532–537, (1994).
8. Dinelli A, Bonetti M, Minelli I and Marotti P, Content of flavonols in Italian bean (*Phaseolus vulgaris* L.) ecotypes, *Food Chem*, 99: 105–114, (2006).
9. Al-Numair KS, Saif Eldein BA, Abdullah HA, Mohammed S and Alamri S, Hydrochloric acid extractable minerals and phytate and polyphenols contents of sprouted faba and white bean cultivars, *Food Chem*, 113: 997–1002, (2009).
10. Ismael DS, Mária T Alena V and Július Á, Influence of cultivar, locality and soil contamination on total polyphenol content and antioxidant activity of Faba bean, *J Micro Bio Food Sci*, 1: 931-941, (2012).
11. Chaieb N, Johannes LG, Montserrat LM, Mohamed B and Manuel V, Polyphenol content and antioxidant capacity of thirteen faba bean (*Vicia faba* L.) genotypes cultivated in Tunisia, *Food Res International*, 44: 970–977, (2011).
12. Baginsky C, Peña-Neira A, Cáceres A, Hernández T, Estrella I, Morales H, Pertuzé R, Phenolic compound composition in immature seeds of fava for raw seeds, green seeds and sprouts of ten Egyptian faba bean cultivars. Significant differences between tested cultivars were detected for total phenolics, tannins and total flavonoids contents as well as for the values of DPPH and FRAP antioxidant activities. The greatest accumulation of all tested phenolic compounds and the highest antioxidant activities occurred in green seeds as compare with either raw seeds or sprouts. In this study it was found that the green faba bean seeds are a good source of bioactive compounds in Egyptian diet with health-promoting antioxidants.

- bean (*Vicia faba* L.) cultivars cultivated in Chile, *J Food Comp Analysis*, 31: 1–6, (2013).
13. Doblado R, Frias J and Vidal-Valverde C, Changes in vitamin C content and antioxidant capacity of raw and germinated cowpea (*Vigna sinensis* var. carilla) seeds induced by high pressure treatment, *Food Chem*, 101:918–923, (2007).
 14. Cevallos-Casals BA and Cisneros-Zevallos L, Impact of germination on phenolic content and antioxidant activity of 13 edible seed species, *Food Chem*, 119: 1485–1490, (2010).
 15. Singleton VL and Rossi JA, Colorimetry of total phenolics with phosphomolybdic – phosphtungstic acid reagents, *Amer J Enol Viticu*, 16: 144-158, (1965).
 16. Osoro K, Mateos-Sanz A, Frutos P, Garcí'a U, Ortega-Mora L M, Ferreira L M M, Celaya R and Ferre I, Anthelmintic and nutritional effects of heather supplementation on Cashmere goats grazing perennial ryegrass-white clover pastures, *J Animal Sci*, 85: 861–870, (2007).
 17. Zhishen J, Mengcheng T and Jianming W, The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals, *Food Chem*, 64: 555-559, (1999).
 18. Brand-Williams W, Cuvelier ME and Berset C, Use of free radical method to evaluate antioxidant activity, *Lebensmittel Wissenschaft Und Technol*, 28: 25-30, (1995).
 19. Benzie IFF and Strain JJ, Ferric reducing/antioxidant power assay: Direct measure of total antioxidant activity of biological fluids and modified version for simultaneous measurement of total antioxidant power and ascorbic acid concentration, *Methods Enzym*, 299: 15–27, (1999).
 20. McKeehen J D, Busch RH and Fulcher RG, Evaluation of wheat (*Triticum aestivum* L.) phenolic acids during grain development and their contribution to fusarium resistance, *J Agri Food Chem*, 47: 1476-1482, (1999).
 21. Lin PY and Lai HM, Bioactive compounds in legumes and their germinated products, *J Agri Food Chem*, 54: 3807–3814, (2006).
 22. Lopez-Amoros ML, Hern´andez T and Estrella I, Effect of germination on
 23. legume phenolic compounds and their antioxidant activity, *J Food Compos Anal*, 19:277–283, (2006).
 24. Sreeramulu D, Vijaya K, Reddy C and Raghunath M, Antioxidant activity of consumed cereals, millets, pulses and legumes in India, *Indian J Biochem Biophys*, 46:112–115, (2009).
 25. Nozzolillo C, Ricciardi L and Lattanzio V, Flavonoid constituents of seed coats of *Vicia faba* (Fabaceae) in relation to genetic control of their color, *Can J Botany*, 67: 1600–1604, (1989).
 26. Tomá's-Barbera'n FA, Garcia-Grau MM and Tomas-Lorente F, Flavonoid concentration changes in maturing broad bean pods, *J Agric Food Chem* 39:255–258, (1991).
 27. Duenas M, Hernandez T, Estrella I and Fernandez D, Germination as a process to increase the polyphenol content and antioxidant activity of lupin seeds (*Lupinus angustifolius* L.), *Food Chem*, 117: 599-607, (2009).